

1 **REVIEW PAPER**

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3 **Use of nociceptive threshold testing in cats in experimental and clinical settings: a**  
4 **qualitative review**

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17 **Running head:** Nociceptive threshold testing in cats

18

19 **Roles and responsibilities for this manuscript:**

20 Chiara Adami: Study design, literature search, manuscript preparation and revision

21 Claudia Spadavecchia: Contribution to the design and critical revision of the manuscript

22

23 **Conflicts of interest:** The authors declare no conflict of interest.

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Journal Pre-proof

27 **Abstract**

28

29 **Objectives** The objective of this work was to review the scientific articles on the use of  
30 nociceptive threshold testing (NTT) in cats, and to summarise the clinical and  
31 experimental applications in this species.

32

33 **Databases used** Pertinent literature was searched with PubMed, Scopus, Web of  
34 Science, Universitätsbibliothek Basel (swissbib Basel Bern) and Google Scholar. The  
35 search was then refined manually based first on article titles and abstracts, and  
36 subsequently on full texts.

37

38 **Conclusions** Of the four classical acute nociceptive models used for NTT, thermal and  
39 mechanical are most commonly used in cats. Thermal stimulation is applicable in  
40 experimental settings and has been used in pharmacodynamics studies assessing feline  
41 antinociception. Although mechanical stimulation is currently less used in cats, in the  
42 future it might play a role in the evaluation of clinical feline pain. However, the low  
43 response-reliability after stimulus repetition within a narrow time interval represents a  
44 major limitation for the clinical use of mechanical thresholds (MT) in this species.

45 Challenges remain when thermal thresholds (TT) are used to investigate  
46 analgesics that have the potential to affect skin temperature, such as opioids and alpha  
47 2-adrenergic agonists, and when a model of inflammatory pain is reproduced in  
48 experimental cats with the purpose of evaluating NSAIDs as analgesics.

49

50 **Keywords** feline, mechanical and thermal thresholds, nociceptive model, nociceptive  
51 threshold testing, pain

## 52 **Introduction**

53 Nociceptive threshold testing (NTT) makes use of a broad range of stimulation methods  
54 to assess and quantify nociceptive function and response. Most protocols described in  
55 cats have been developed for cutaneous application of mechanical and thermal  
56 stimulation. However, mechanical NTT has also been successfully applied to hollow  
57 viscera (Briggs et al. 1998).

58       Regardless of the type of stimulation used in the experimental setting, a realistic  
59 reproduction of clinical pain is probably impossible to achieve. Although characterised  
60 by a number of different features, a common denominator of clinical pain is its  
61 complexity and the diversity of the nociceptors involved where mechanical, thermal and  
62 chemical stimuli may all contribute to the activation of afferent pathways during  
63 postoperative surgical pain.

64       Nociception and pain are considered distinct processes. Nociception begins with  
65 the detection of injurious stimuli by a class of specialised receptors, with transmission  
66 of that information to the spinal cord and on up to the brain. This may result in a  
67 defensive, immediate reflex response (Sneddon, 2017). All reflexes, including those  
68 associated with nociception, are organised by centres at the lower hierarchy of the  
69 central nervous system; they can be elicited in decerebrated animals and are  
70 characterised by either autonomic or basic motor responses, including increased heart  
71 rate, withdrawal and muscular contractions (Woodworth & Sherrington 1904; Sneddon  
72 2017). Complex behaviours, in response to noxious stimuli, can also include  
73 conditioned motor responses, usually as a result of learning (Le Bars et al. 2001). Pain  
74 is a negative affective and psychological response and is often accompanied by more  
75 complex or prolonged behavioural alterations indicative of discomfort, such as distrust  
76 of objects associated

77 with painful experiences and/or modification of social behaviour. Nociception may not  
78 result in pain, because of the ability of the central nervous system to modify nociceptive  
79 signals and prevent conscious perception of noxious stimuli, and pain can occur without  
80 nociception in the presence of central sensitisation. However, pain resulting from injury  
81 cannot occur without nociception (Sneddon 2017).

82       There is a huge body of literature on NTT in rodents, however these animals are  
83 often genetically very similar, leading to minimal variance in the detected thresholds.  
84 Cats used in research are, by comparison, much more genetically diverse and so it is  
85 expected that there will be greater variance in any tested population. There are many  
86 reports focusing on the application of NTT to cats. Older reports were aimed at  
87 investigating particular aspects of the afferent nociceptive organisation, or at  
88 establishing patterns to relate the neurophysiological activity of the sensory system to  
89 behavioural responses indicative of nociception (Beck et al. 1974; Casey & Morrow  
90 1983). In contrast, more recent work has primarily focused on the pharmacodynamics of  
91 analgesic drugs in cats, with the purpose of identifying useful doses, routes of  
92 administration, onset times and duration of the effects (Millette et al. 2008; Pypendop et  
93 al. 2009; Ambros & Duke 2013).

94       When performing NTT testing it is important for the stimulus to be applied at a  
95 rate that will allow for conduction and interpretation of the stimulus. If the increase in  
96 stimulus strength is very rapid it might reach an excessively high level before the  
97 animal has had a chance to respond. It is also important that the approach can detect  
98 hyperalgesia as well as analgesia. In tests that use latency to the response, a very short  
99 latency will not likely allow for the detection of hyperalgesia.

100 The objective of this work is to review the use and the clinical and experimental  
101 applications of NTT in cats, with particular focus on acute nociceptive models, and on  
102 the literature of the past 20 years.

103

#### 104 **Databases used and literature search**

105 A literature search was conducted using PubMed, Scopus, Web of Science,  
106 Universitätsbibliothek Basel (swissbib Basel Bern) and Google Scholar. The keywords  
107 sets used for the initial screening were the following: ‘nociceptive threshold testing +  
108 cats/feline’, ‘quantitative sensory testing/QST + cats/feline’,  
109 ‘mechanical/thermal/electrical thresholds + cats/feline’, ‘mechanical/thermal/electrical  
110 nociceptive model/antinociception + cats/feline’, ‘antinociceptive/analgesic  
111 effects/efficacy + cats/feline’, and ‘antinociceptive/analgesic pharmacodynamics +  
112 cats/feline’.

113 The search was refined based first on article titles and abstracts, and subsequently  
114 on full texts of the selected scientific reports. The reference list of each retrieved  
115 scientific paper was then scrutinised to identify further pieces of literature pertaining to  
116 the topic. All the identified scientific peer-review articles written in the English  
117 language and pertaining to the topic were included in the study. Related anaesthesia and  
118 neurophysiology textbooks were also reviewed.

119 The refined search identified 51 articles published between 1983 and 2019. Of  
120 these, nine were on the use of mechanical thresholds (MT) (of which six were  
121 experimental and three were clinical studies), 34 experimental studies were on the use  
122 of TTs, and eight on the use of both mechanical and TTs (of which one was a clinical  
123 report and the remaining seven were experimental studies).

124

**125 Mechanical stimulation**

126 Mechanical stimulation has been used in cats to elicit mostly somatic, but also visceral  
127 nociception, in both experimental and clinical settings. Visceral nociception has been  
128 experimentally induced by inflating balloon catheters inserted into the rectum (Sawyer  
129 & Rech 1987; Briggs et al. 1998). Somatic nociception is induced and assessed by  
130 applying a force to a given area of the body. Force is defined as the push or pull on an  
131 object that causes it to change velocity; pressure is a measure of force per unit area.  
132 Therefore, for the outcome values to be comparable between devices that measure  
133 different variables, the surface area of the probe must be known, and recognised as part  
134 of the applied stimulus. A nociceptive threshold is defined, depending on the device  
135 used, as either the pressure (expressed in mmHg) or the force (expressed in g or  
136 Newtons) reached when the stimulus is intense enough to elicit a behavioural, conscious  
137 response in the cat, which is subjectively determined by the operator. Ideally, the  
138 stimulating probe should be applied perpendicular to the test site in order to ensure that  
139 the measured force has been wholly applied to the area of interest. There should be  
140 minimal distensible tissue so that the mechanical stimulus is not spread over a large  
141 surface area or that the stretching tissue attenuates the force applied. Mechanical stimuli  
142 are usually applied progressively and incrementally until a cut-off value is reached; the  
143 speed of increased force is variable and, for manual algometers, is operator dependent.  
144 Both sharp-ended pins and flat-ended/blunt probes have been used in animals (Moens et  
145 al. 2003; Haussler & Erb 2006; Machin et al. 2019).

146 It is commonly accepted that the elicited behaviours represent supraspinal  
147 responses to activation of nociceptors located in the skin, muscles and periosteum (Le  
148 Bars et al. 2001). Both myelinated A $\delta$  fibres, with intermediate conduction speeds, and  
149 small, unmyelinated, slow-conducting C fibres are expected to be activated primarily,

150 but conventional noxious mechanical stimuli do not produce selective activation of  
151 these nerve types (Le Bars et al. 2001). In some subjects/patients, activation of the A $\beta$   
152 fibres, in response to touch and pressure, may be sufficient to evoke behavioural  
153 responses that could easily be misinterpreted as signs of nociception. This is more likely  
154 in the case of algometers that are applied intermittently than those that maintain contact  
155 with the skin where the A $\beta$  stimulation would be ongoing. As a result, one disadvantage  
156 of intermittently applied mechanical stimulation is a potential lack of specificity.

157 Several instruments have been specifically developed to perform mechanical  
158 stimulation in cats (Table 1). Slingsby and colleagues designed a finger-mounted  
159 algometer (Slingsby et al. 2001). The probe was made of a Force-Sensing Resistor  
160 (FSR), a thick polymer film, which exhibits a progressive decrease in resistance with  
161 increasing force applied to its surface. The 15 mm diameter-probe was soldered to a  
162 ribbon cable, connected to a battery powered measuring unit, calibrated with an accurate  
163 load beam and mounted on the index finger of the operator. The outcome force resulting  
164 from the application of the probe on the skin of the cat was expressed in Newtons. This  
165 device was subsequently used to evaluate the analgesic effect of post-operative  
166 meperidine in male cats undergoing castration, and in another clinical study  
167 investigating NSAID associated analgesia in 40 female cats undergoing  
168 ovariohysterectomy (OVH) (Slingsby & Waterman-Pearson 2000; Slingsby et al. 2001).  
169 Changes in MTs measured at the scrotum before and after surgery differentiated  
170 between meperidine and the negative control group in male cats. In female cats,  
171 thresholds measured at the surgical wound following OVH were lower than those  
172 measured before surgery. The authors did not state how quickly the force was applied in  
173 either study and did not describe how or where the probe was applied in the OVH study  
174 (Slingsby & Waterman-Pearson 2000; Slingsby et al. 2001).

175 Another pressure testing device for use in cats was designed in 2007 by Dixon and  
176 colleagues (Dixon et al. 2007) and subsequently manufactured by Topcat Metrology Ltd  
177 (UK), marketed under the trade name “ProD-Plus”. The device was composed of a 5 g  
178 plastic bracelet, inside which the authors mounted a blood pressure bladder and three  
179 brass pins, each tipped with a 2.4 mm diameter ball bearing, distributed 10 mm apart in  
180 a triangular pattern to apply a perpendicular pressure to the limb. For this device, the  
181 outcome was bladder pressure expressed in mmHg. This could not be translated into the  
182 force acting on the skin because neither the true contact area nor the actual pressure  
183 applied to each pin was known. The bracelet was applied on one forearm and bladder  
184 pressure was increased incrementally and measured with a strain gauge pressure  
185 transducer; the threshold pressure was recorded by pressing the hold button on the  
186 voltmeter when the cat reacted to the stimulus (the voltage was directly proportional to  
187 the pressure). The authors found that, whilst the thresholds varied a lot between  
188 different cats (68 to 202 mmHg in six cats), thresholds within each cat were consistent.  
189 This pressure algometer was used by the same authors to evaluate the analgesic efficacy  
190 of subcutaneous butorphanol ( $0.4 \text{ mg kg}^{-1}$ ) and carprofen ( $4 \text{ mg kg}^{-1}$ ). The NSAID was  
191 tested in a second phase of the trial, after kaolin was injected intradermally in the  
192 forearm to produce a model of inflammatory pain. In that study, the comparison  
193 between MTs measured before and after the administration of the opioid detected  
194 butorphanol antinociception. Excessively variable thresholds were obtained with the  
195 inflammatory model making this approach ineffective. The authors concluded that the  
196 device was light and easy to use and allowed the cats to remain unrestrained during  
197 testing. The repeatability of the thresholds was considered acceptable by the authors,  
198 who concluded that the algometer could be used for analgesic pharmacologic studies in  
199 cats (Dixon et al. 2007). However, the same authors developed and tested the device,

200 which may have resulted in a certain degree of bias, although the device has been  
201 further modified and used by others who have confirmed its utility (Steagall et al. 2007;  
202 Millette et al. 2008; Slingsby et al. 2012).

203 Mechanical thresholds have also been used in cats to study experimentally  
204 induced visceral nociception (Sawyer & Rech 1987; Sawyer et al. 1993). A subsequent  
205 study (Briggs et al. 1998) investigated the analgesic effect of oxymorphone,  
206 butorphanol and acepromazine, alone and in combination, using this model. A silastic  
207 balloon catheter, inserted into the rectum and connected via a rubber tube to a plastic  
208 jug was pressurised to selected incremental values for 30-second periods. A positive  
209 result was considered when an undefined behavioural response was evoked. The authors  
210 interspersed lower pressures to prevent conditioning to the increasing pressures used.  
211 Although the model has been successfully used in horses (Muir & Robertson 1985) and  
212 found robust and reliable in rodents (Jones et al. 2004; Arvidsson et al. 2006; Nissen et  
213 al. 2018), its validity for investigating colorectal noxious distension was questioned by a  
214 more recent study in horses (Sanchez et al. 2005). The physical properties of the balloon  
215 are relevant and materials with linear compliance, such as mylar, should be selected  
216 over latex in order to ensure proper pressure application to the colorectal wall (Sanchez  
217 et al. 2005). Another drawback of this model is that it may fail to differentiate  
218 behavioural responses caused by nociception from those caused by an urge to defecate.

219 More recent studies used precision pressure algometers, such as the Electronic  
220 von Frey Anaesthesiometer (EVF) and the Small Animal ALGOmeter (SMALGO). The  
221 former represents the electronic version of the von Frey filaments and has been used  
222 most recently to assess acute and chronic pain in dogs and cats (Adami et al. 2018;  
223 Addison & Clements 2017). The EVF uses a sensory probe equipped with a rigid tip  
224 applying a force varying from 0 to 1000 g, which is measured, displayed and stored by

225 the control unit. The SMALGO has been specifically developed for laboratory rodents  
226 and shares the same working principle as the EVF. However, it has a finger-mounted  
227 sensor probe whose applicable force ranges from 0 to 1500 g. With both devices, a  
228 progressively increasing force is applied by the operator over the targeted area, until an  
229 end-point behavioural response is observed. Although the stimulus is generally applied  
230 over an undefined time-period, most authors set a cut-off pressure/force value to avoid  
231 iatrogenic injury.

232 Addison and Clements (2017) found that use of both the von Frey filaments and  
233 the EVF, applied to the metacarpal/metatarsal pad to assess paw withdrawal thresholds,  
234 resulted in differentiation between healthy cats and those with osteoarthritis (Addison  
235 and Clements 2017). Another recent study evaluated the inter-rater and inter-device  
236 reliability of TTs measured with both the EVF and the SMALGO in non-painful cats  
237 (Adami et al. 2018). The authors found that the reliability of the measurements  
238 decreased after repetition within time-intervals shorter than one hour, indicating that the  
239 level of cooperation of feline patients may decrease after repeated testing or,  
240 alternatively, that the cats may anticipate the stimulus in order to end it. Similarly,  
241 learning and stimulus anticipation, resulting in decreased TTs have been described in  
242 dogs using algometry (Coleman et al. 2014). In another study, the SMALGO, applied to  
243 the skin of the upper lip, dorsal to the end of the canine root, was evaluated in cats with  
244 chronic gingivostomatitis, as compared to a healthy control group (Machin et al.  
245 2019b). Although the authors found a low inter-observer and intra-observer variability,  
246 the study failed to differentiate between healthy and diseased cats. Moreover, there was  
247 no correlation between the scores of the chronic gingivostomatitis scale, used by the  
248 authors to score the severity of the oral lesions, and the thresholds measured in diseased

249 cats. Overall, these findings suggested that mechanical sensory testing with the  
250 SMALGO is not a reliable method to evaluate chronic oral pain in cats.

251 A final study compared the use of the EVF and von Frey filaments at different  
252 anatomical sites of non-painful cats (Machin et al. 2019a). The authors found a  
253 moderate agreement between the two devices, as suggested by the intra-class correlation  
254 coefficient of 0.49 (CI=0.13-0.70); however, the willingness of the cats to cooperate  
255 decreased with the repetition of the measurements after 24 hours. This drawback may  
256 limit the applicability of mechanical NTT in the clinical setting, where repeated testing  
257 may be needed to adjust the analgesic therapy to meet the patient's requirement. Despite  
258 its limitations, it is worth considering that one potential advantage of the mechanical  
259 NTT over thermal, electrical and chemical techniques may be that the use of pressure  
260 thresholds is often perceived, by both clinicians and cat owners, as less invasive and  
261 harmful than other types of stimuli. This aspect may allow and encourage the  
262 development of protocols to increase the clinical application of TT testing, for routine  
263 assessment of pain in cats.

264 At the date of writing, although inter-observer variability appears to be minimal  
265 for non-repeated TT measurements, there is still no evidence that, in cats, the thresholds  
266 consistently correlate with the severity of the clinical condition that causes pain, or with  
267 the intensity of pain itself. More prospective studies on the use of TTs in cats with  
268 clinical pain are needed to draw more solid conclusions with respect to the clinical  
269 usefulness of modern pressure algometers in this species. Even when used for testing  
270 analgesics in an experimental setting the documented thresholds have been highly  
271 variable making it difficult to record an antinociceptive effect.

272

273 **Thermal stimulation**

274 A number of studies investigated the applicability of TT testing in various feline  
275 nociceptive models (Table 2). Thermal stimulation relies on the activation of two  
276 subtypes of nociceptors: mechano-heat units activated by noxious mechanical and hot  
277 thermal stimuli, and mechano-cold units activated by noxious mechanical and cold  
278 thermal stimuli (Djoughri & Lawson 2004). Two different outcome variables are often  
279 used during thermal stimulation: latency (the time elapsed between the start of  
280 application of a constant temperature and the observation of the target behavioural  
281 response) and threshold temperature (the measured temperature at which the response  
282 occurs with the application of an increasing temperature) (Casey & Morrow 1983;  
283 Slingsby & Taylor 2007; Addison & Clements 2017). Hot thermal testing is most  
284 commonly described, although cold (7°C temperature-controlled pressure-plate system)  
285 has also been used in cats and found more useful than kinetic gait analysis to  
286 differentiate between healthy limbs and those with osteoarthritis (Addison & Clements  
287 2017).

288         There is a general concern that noxious thermal stimulation may activate mostly C  
289 but not A $\delta$  fibres and therefore result in incomplete activation of nociceptive pathways,  
290 making the thermal model less likely than others to resemble the complexity of clinical  
291 pain (Mao 2012). Selectivity of receptor activation is greatly dependent on the mode of  
292 delivery of the thermal stimulus and on the steepness of the heating slope. In a murine  
293 model, stimuli capable of heating the cutaneous surface as rapidly as 6.5°C second<sup>-1</sup>,  
294 such as laser radiation, activated A $\delta$  units with a response latency of 2 seconds after the  
295 onset of the stimulus (Yeomans & Proudfit 1996). In contrast, thermal conduction, by  
296 means of rates of skin heating as slow as 0.9 °C second<sup>-1</sup>, with relatively long latency of  
297 5-6 seconds, evoked action potentials selectively in C nociceptors (Yeomans & Proudfit  
298 1996). In cats, as well as in primates, thermal stimuli above 45 °C are capable of

299 activating both the A $\delta$  and C fibres, which respond with increasing discharge as the  
300 temperature is increased (Beck et al. 1974; Casey & Morrow 1983). Both laser and  
301 radiant heat stimulation responses are measured as latencies, whereas temperature  
302 thresholds are measured with contact thermodes. Contact thermodes unavoidably apply  
303 a pressure on the skin surface, which may also activate low-threshold non-nociceptive  
304 A $\beta$  fibres (Nathan et al. 1986; Svensson et al. 1997) but this is less likely to cause  
305 confusion if the thermode is continuously in contact with the skin vs an intermittent  
306 application.

307         Despite these limitations, thermal nociception has been used extensively over the  
308 last two decades and its use in cats has been more repeatable and reliable than both  
309 mechanical and electrical models. The use of cats to investigate and quantify afferent  
310 activity in response to thermal stimulation dates back to the 1960s (Kenshalo et al.  
311 1967; Beck et al. 1974). One study used rapid onset thermal pulses ranging from 43 to  
312 60°C and concluded that the probability of evoking a nocifensive response in cats  
313 increased for cutaneous thermal stimuli between 50 and 55°C (Casey & Morrow 1983).

314         In 2002, Dixon and colleagues developed a TT device, subsequently produced by  
315 Top Cat Metrology Ltd, which has been used for evaluation of various analgesics in cats  
316 (Dixon et al. 2002). A probe equipped with a heater element and a temperature sensor  
317 was held against a clipped area of the thorax using an elastic band and a pressure  
318 bladder, inflated to 100 mmHg to ensure even contact between the skin and the probe.  
319 The probe was heated at 0.6°C second<sup>-1</sup> until either a pre-defined behavioural response  
320 was elicited, or a cut-off value of 60°C was reached. The measurement of the TTs with  
321 this device was repeatable and well tolerated by the cats but resulted in minor skin  
322 lesions. A further crossover trial carried out by the same authors in non-painful cats

323 found the device useful to differentiate between pethidine antinociception and a placebo  
324 treatment (Dixon et al. 2002).

325 This device was used in various subsequent studies with a standardised approach,  
326 characterised by a cut-off temperature decreased to 55-55.5°C, to prevent skin lesions,  
327 the same application mode of the thermal stimulus and similar areas of the body tested.  
328 Many of these reports investigated the usefulness of TTs alone to evaluate the  
329 pharmacodynamics of various analgesics (Lascelles & Robertson 2004b; Robertson et  
330 al. 2005; Johnson et al. 2007; Wegner & Robertson 2007; Slingsby & Taylor 2008;  
331 Robertson et al. 2009; Slingsby et al. 2009; Slingsby et al. 2010) (Table 2), whereas  
332 some others compared mechanical and TT testing for the same purpose (Steagall et al.  
333 2006; Steagall et al. 2007; Millette et al. 2008; Slingsby et al. 2012; Ambros and Duke  
334 2013; Addison & Clements 2017) (Table 3).

335 In the context of pharmacological studies, TT testing has been commonly used to  
336 describe the analgesic effects of various opioids in cats, including “opioid-like” agents  
337 such as tramadol and tapentadol (Lascelles & Robertson 2004a; Johnson et al. 2007;  
338 Wenger & Robertson 2007; Pypendop et al. 2009; Steagall et al. 2015; Doodnaught et  
339 al. 2017). Overall, the results of these studies suggest that the thermal nociceptive  
340 model consistently detects opioid-antinociception, despite some contradictory findings;  
341 whilst buccal buprenorphine was found by some authors to significantly increase TTs  
342 (Robertson et al. 2005a; Doodnaught et al. 2018), it resulted in inconsistent thermal  
343 antinociception in another study (Steagall et al. 2015). A possible reason for these  
344 conflicting results is the variable bioavailability of buprenorphine after oral  
345 transmucosal administration, which was found to range between 16 and 60% (Pypendop  
346 et al. 2014).

347 An important drawback of using TTs to detect opioid antinociception in cats is  
348 that opioids increase body temperature (Niedfeldt & Robertson 2006; Posner et al.  
349 2010), an effect that may act as a confounding variable and potentially affect the  
350 outcome because the raised baseline temperature may not be comparable to an untreated  
351 control.

352 The thermal nociceptive model has also been used to investigate the analgesic  
353 pharmacodynamics of  $\alpha_2$ -adrenoceptor agonists in cats, with conflicting results.  
354 Slingsby & Taylor (2008) found that, among five different intramuscular  
355 dexmedetomidine doses tested, only the highest one ( $40 \mu\text{g kg}^{-1}$ ) caused an increase in  
356 thresholds, which was less significant than with buprenorphine, used as positive control  
357 treatment. Another study failed to detect any difference using TT between intramuscular  
358 and oral transmucosal dexmedetomidine at the same dose (Slingsby et al. 2009).

359 In a subsequent report the same authors detected an additive antinociceptive effect  
360 with buprenorphine and dexmedetomidine combined in non-painful cats (Slingsby et al.  
361 2010). In the light of these inconsistent findings, the authors concluded that the  $\alpha_2$ -  
362 adrenoceptor agonists-induced vasoconstriction may alter the response to thermal  
363 stimulation by decreasing blood flow in the skin, which makes the thermal model  
364 suboptimal when this class of analgesics is investigated (Slingsby et al. 2009). The  
365 decrease in skin temperature appears to be dose dependent with doses  $>5 \mu\text{g/kg}$  causing  
366 some decrease (Pypendop personal communication 2020).

367 Besides the contact thermal algometer developed by Dixon (Dixon et al. 2002),  
368 another device using remote carbon dioxide laser stimulation has been validated for cats  
369 more recently (Farnworth et al. 2013a, Farnworth et al. 2013b). A visible, non-thermal,  
370 helium laser was used to guide and aim the thermal carbon dioxide laser beam over a  
371 target area of the cats' shaved thorax. The wavelength of the thermal laser was  $10.6 \mu\text{m}$ ,

372 with a maximum power output of 10 W. The exhibition of either a body shift (e.g.  
373 rising) or the panniculus reflex were considered a positive response. The laser device  
374 was evaluated with respect to intra-individual and inter-individual variability. The  
375 authors found that, although individual responses were repeatable over a three-day  
376 period, the repeatability decreased after the third day of testing (Farnworth et al. 2013a).  
377 Moreover, heavier cats had increased latencies, suggesting that fat deposition in the sub-  
378 cutaneous layers, where the skin nociceptors occur, may act as a buffer and attenuate the  
379 response (Farnworth et al. 2013b). This laser device was used to investigate the  
380 analgesic effects of opioids, NSAIDs and  $\alpha_2$ -adrenoceptor agonists (Farnworth et al.  
381 2015). Although the results of this one study were inconclusive and did not allow  
382 differentiation between treatment groups, it is worth considering that using a CO<sub>2</sub> laser  
383 thermal stimulator may offer some advantages over other types of thermal probes. The  
384 monochromatic, long wavelength results in complete absorption regardless of the  
385 degree of pigmentation of the skin, which may be an issue with radiant heat methods  
386 (Le Bars et al. 2001). Moreover, the heating slope is steeper than with contact  
387 thermodes as the target temperature is reached within milliseconds, and the lack of  
388 cutaneous contact ensures avoidance of inadvertent activation of non-nociceptive nerve  
389 fibres (Treede et al. 1984; Le Bars et al. 2001). However, one potential disadvantage is  
390 the lack of skin temperature measurement before application of the stimulus, as well as  
391 the risk for blistering that was observed in some study cats.

392 Overall, many studies suggest that thermal nociception is well tolerated in cats  
393 and the results are reproducible (Lascelles & Robertson 2004a; Robertson et al. 2005a;  
394 Robertson et al. 2005b; Steagall et al. 2006; Pypendop et al. 2009). A limitation of TTs  
395 may be their applicability to pharmacodynamic studies focusing on analgesics that have  
396 the potential to alter the skin temperature in cats, such as opioids (Niedfeldt &

397 Robertson 2006; Posner et al. 2007) and  $\alpha_2$ -adrenoceptor agonists at high doses - an  
398 effect that may affect the thresholds or the comparison with control animals and act as a  
399 confounding variable. The thermode method is unlikely to be very useful for  
400 investigating pain in clinical patients because it is time consuming, requires a skin area  
401 to be shaved for its placement and the repeated application of noxious heat may be  
402 regarded as upsetting by some animal caregivers. Methods that are more 'portable',  
403 such as a laser, that can be focused on an area of interest and used intermittently, may  
404 be more useful in the clinical setting but may still be unacceptable due to the repeated  
405 testing (Farnworth et al. 2015).

406

#### 407 **Electrical stimulation**

408 The potential advantage of electrical stimulation is that it is reproducible, measurable  
409 and quantifiable. Single electrical stimuli of short duration, usually between 10 and 20  
410 ms, are often applied in a sudden, abrupt fashion to measure latencies. Alternatively,  
411 electrical stimulation of gradually increasing intensity in the form of trains of stimuli,  
412 usually lasting some hundreds of ms, have been used in rodents to evaluate different  
413 responses organised on a hierarchical basis, namely reflex movements of the tail  
414 followed by more complex behavioural responses, such as attempts to escape (Le Bars  
415 et al. 2001).

416 A number of studies from the 1970s described the use of electrical stimulation in  
417 cats (Anderson & Pearl 1974; Berkley & Parmer 1974; Anderson & Pearl 1975; Lisney  
418 1978). However, the majority of these neurophysiological studies were conducted in  
419 cats under general anaesthesia and did not use behavioural evaluation, which is an  
420 intrinsic component of NTT.

421 More recently, Millette and colleagues evaluated the use of electrical threshold  
422 testing to characterise the antinociceptive effects of meperidine in pain-free cats  
423 (Millette et al. 2008). The authors used a current generator to deliver repeated stimuli  
424 with a duration of 1 ms and 1 ms delay between pulses, through two electrodes held  
425 against a clipped area of the mid-thorax. The current was increased at 1mA second<sup>-1</sup>,  
426 and the cut-off was set at 5 mA. The main finding of Millette's study was that the  
427 electrical stimulus failed to detect meperidine antinociception, whereas the thermal and  
428 the TTs, also used by the authors, were both found useful for this purpose.

429 The application of electrical stimulation for nociceptive testing in cats has failed  
430 to earn popularity. A reason for this may be the many limitations of the electrical  
431 stimulus, which differs from every natural type of stimulus that an animal may  
432 encounter in its physiological environment.

433 Although studies conducted in both human volunteers and dogs demonstrated that  
434 electrical stimuli with frequencies of 2000, 250 and 5 Hz can selectively stimulate the  
435 A $\beta$ , the A $\delta$  and the C fibres, respectively (Finkel et al. 2002; Sakai et al. 2004; Watabiki  
436 et al. 2010), to the best of these authors' knowledge, there are no published experiments  
437 in cats to verify these findings. As a result, nonselectivity of activation is another  
438 potential drawback of electrical stimulation, which can result in activation of A $\delta$ , C as  
439 well as larger diameter fibres not directly implicated in nociception (Le Bars et al.  
440 2001). Finally, there are technical considerations that may limit the applicability of  
441 electrical nociception. Based on its thickness and hydration, the skin offers variable  
442 impedance to electrical stimulation, which can considerably affect the response. This  
443 variability can be minimised by using a constant current and measuring impedance prior  
444 to stimulation (Le Bars et al. 2001).

445           The very limited evidence, together with the small number of reports in this  
446 species, does not allow any conclusive statement with respect to the usefulness of the  
447 electrical nociceptive model in cats.

448

#### 449 **Chemical stimulation**

450 Chemical stimuli differ from any other type of nociceptive stimulation, as they are slow,  
451 progressive and of longer duration. As a result, the chemical nociceptive model mostly  
452 produces complex behavioural responses rather than simple reflexes (Le Bars et al.  
453 2001). Algogenic or irritant agents, such as capsaicin, formalin and kaolin, are either  
454 applied on the intact skin or injected subcutaneously or intradermally, to produce  
455 hyperalgesia and inflammation, and therefore evoke pain. A local cutaneous injury may  
456 produce primary hyperalgesia within the injured area, as well as secondary, neurogenic  
457 hyperalgesia, caused by central sensitisation, in the normal surrounding skin. (Baumann  
458 et al. 1991).

459           The duration of inflammation – and therefore of hyperalgesia – varies between  
460 chemical agents, routes of administration and, possibly, animal models. In rodents, both  
461 formalin and capsaicin reproduce an inflammatory model characterised by two well-  
462 identified phases, of which the acute one occurs within minutes from intradermal  
463 injection and lasts a few minutes, followed by secondary hyperalgesia starting around  
464 10 minutes (Wheeler-Aceto & Cowan 1991; La et al. 2017). In cats, subcutaneous  
465 injection of kaolin in the paw results in well-defined and reproducible inflammation that  
466 lasted up to five days (Giraudel et al. 2005; Giraudel et al. 2009).

467           The failure to quantify NSAID associated analgesia and successfully differentiate  
468 between different agents within the same pharmaceutical class seems to be a common  
469 denominator of the studies that used the mechanical and thermal nociceptive models.

470 Conceptually, since these models do not have an inflammatory component to the  
471 nociceptive stimulus it is not surprising that they have not succeeded. The addition of an  
472 inflammatory chemical has been used to test the anti-inflammatory antinociceptive  
473 effect of these drugs.

474 A study from the late 1970s used formalin, injected subcutaneously into the  
475 forepaw, to induce inflammation and then quantify the analgesic effects of morphine  
476 and meperidine with thermal latency testing in cats (Dubuisson & Dennis 1977).  
477 Similarly, kaolin was used in various studies to investigate NSAID associated  
478 antinociception in cats (Giraudel et al. 2005; Dixon et al. 2007; Taylor et al. 2007;  
479 Giraudel et al. 2009). Taylor and colleagues combined kaolin injection and TT testing to  
480 investigate the analgesic efficacy of ketoprofen in seven cats, and found that the  
481 combination of the two techniques did not detect antinociception. Ketoprofen increased  
482 the TT outside the 95% confidence interval but the study was probably underpowered  
483 (Taylor et al. 2007).

484 The intradermal or subcutaneous injection of chemicals has been used only in  
485 experimental cats with the greatest utility for testing the analgesic effect of drugs that  
486 have an anti-inflammatory component. There are no potential applications in the clinical  
487 setting.

488

#### 489 **Mixed nociceptive models comparing thermal and mechanical thresholds**

490 A number of studies conducted in cats reported the simultaneous use of several  
491 threshold testing methods to investigate the pharmacodynamics of various analgesic  
492 agents, most of which were opioids (Steagall et al. 2006; Steagall et al. 2007; Millette et  
493 al. 2008; Steagall et al. 2008; Slingsby et al. 2012; Table 3).

494 The studies using different types of nociceptive stimulation seem to further  
495 confirm that the TTs, and to some extent the mechanical ones, consistently detect the  
496 antinociceptive effect of opioids (Millette et al. 2008; Slingsby et al. 2012), whilst less  
497 convincing findings were obtained when the two models were used to investigate  
498 ketamine (Ambros & Duke 2013). The authors concluded that, because ketamine seems  
499 to be more effective in pathological pain states characterised by central facilitation  
500 (Ghorpade and Advokat 1994), a model of acute nociception may not be the most  
501 appropriate one to detect ketamine analgesia (Ambros & Duke 2013).

502 Tramadol was found to have a limited effect on both TT and TTs (Steagall et al.  
503 2008), although the increase in thresholds was more pronounced when tramadol was  
504 combined with 0.1 mg kg<sup>-1</sup> acepromazine. However, the sedative effect, detected in all  
505 cats that received acepromazine (Steagall et al. 2008), could represent a confounding  
506 variable, by decreasing the behavioural responsiveness of the cats to nociceptive  
507 stimulation.

508

## 509 **Conclusions**

510 Mechanical and TT testing are the NTT methods that were found more reliable for use  
511 in cats within the last two decades, with TTs being the most widely applied in  
512 pharmacological studies. As indicated above TT testing may be influenced by changes  
513 in skin temperature associated with particular drugs. With a thermode technique the  
514 baseline temperature is measured but this needs to be accounted for with other methods  
515 where the skin temperature is not recorded automatically.

516 Whilst TT testing is mostly applicable to the experimental setting, there is a  
517 promising, increasing tendency to test the usefulness of MTs in cats with clinical pain.  
518 Therefore, mechanical nociception may, in the future, become part of the routine

519 evaluation of feline patients suffering from various pain syndromes. The low  
520 repeatability of mechanical NTT within short time-intervals, as well as the lack of data  
521 in patients with acute and chronic pain, represent major limitations to its clinical  
522 application. Some studies showed that both MT and TT testing did not detect NSAID  
523 associated analgesia, suggesting that, in order to investigate the efficacy of drugs whose  
524 analgesic effect is mostly based on their anti-inflammatory properties, inflammation  
525 must be produced first.

526

### 527 **Conflict of interest statement**

528 The authors declare no conflict of interest.

529

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532

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**Table 1. Studies investigating the use of mechanical thresholds in both experimental and clinical cats (in chronological order)**

Study (authors, year and publication type)	Number of cats	Type of algometer	Sensitive probe characteristics	Pressure/ force reached (range/ cut off value)	Area tested	Analgesics tested	End-point behavioural response as described by the authors	Baseline thresholds (site and time of measurement)
Briggs et al. 1998 (research paper)	8	Silastic balloon catheter inserted per rectum and connected to a pressurised plastic jug	NA	Approximately 0-50 mmHg (no cut-off)	Rectal mucosa	IV butorphanol, oxymorphone and ACP alone and in combination (saline as negative control group)	Stretching of the hind limbs, abdominal muscular contraction, back arching, changes in breathing pattern	Control thresholds measured before any drug administration
Slingsby & Waterman-Pearson 2000 (research paper)	40	Pressure FSR finger-mounted algometer	15 mm diameter flat surface	0.75-0.95 N (no cut-off)	Surgical wound (OVH)	SC Carprofen, Ketoprofen, Meloxicam and Tolfenamic acid	Flinch away from pressure	At the left flank, before premedication
Slingsby et al. 2001 (research paper)	40	Pressure FSR finger-mounted algometer	15 mm diameter flat surface	0-4 N (no cut-off)	Scrotum	IM meperidine (versus noppethidine as negative control)	Cat pulling away	At the scrotum, before surgery
Dixon et al. 2007 (research paper)	11	ProD Plus pressure algometer	Three pins, each tipped with a 2.4 mm diameter ball – bearing pin	600 mmHg (cut-off)	Forearm	SC Butorphanol and Carprofen	Picking up the leg and shaking it, turning the head towards bracelet, licking/biting bracelet, vocalisation	Forearm, before kaolin injection
Ferreira et al. 2011 (research paper)	8	Two different devices: a C clamp and a mechanical algometer	1-cm <sup>2</sup> circular tip	5 and 20 kg cm <sup>2</sup> (cut-off) for the C clamp and the algometer, respectively	Metacarpus and antebrachium	IV and OTM methadone	Cat turning its head toward the stimulus, moving away from the stimulus, vocalising, or attempting to bite	Control thresholds measured before methadone administration
Porters et al. 2014 (research paper)	6	ProD Plus pressure algometer	4 mm diameter probe	20 N (cut off)	Pectoral muscle (shoulder joint)	Combination of dexmedetomidine and buprenorphine either IM or OTM	Jumping, limb withdrawal, head turning, vocalisation	Control thresholds measured before any drug administration

Adami et al. 2018 (research paper)	13	EVF and SMALGO	0.8 mm diameter rigid tip (EVF) and 3 mm diameter pointed tip (SMALGO)	0-1000 g (EVF) and 0-1500 g (SMALGO)	Lumbosacral joint and medial aspect of the stifle	NA (reliability/repeatability study)	Attempts to escape, tail wiggling, hissing, attempts to bite, aggression, ears back and flat against the head, head turning towards the stimulation site, back muscle contraction and limb withdrawal	NA
Machin et al. 2019a (short communication)	15	EVF and VFF	Probe equipped with 0.8 mm diameter rigid tip (EVF)	0-1000 g (EVF) and 0.008-300 g (VFF)	Upper lip and medial aspect of the stifle	NA (reliability/validation study)	Limb/head withdrawal, head turning, watching the application site, vocalisation, hissing, attempts to bite/scratch	NA
Machin et al. 2019b (research paper)	30 (15 healthy cats and 15 cats with CGS)	SMALGO	3 mm diameter pointed tip	0-1500 g	Upper lip above the canine root	NA (reliability/repeatability study)	Limb/head withdrawal, head turning, watching the application site, vocalisation, hissing, attempts to bite/scratch	NA

Table legend: NA: not applicable; IV: intravenous; ACP: Acepromazine; MT: mechanical thresholds; OVH: Ovariohysterectomy; FSR: Force-Sensing Resistor; IM: intramuscular; SC: subcutaneous; NSAIDs: Non-Steroidal Anti-inflammatory Drugs; EVF: Electronic von Frey Anaesthesiometer; OTM: oral transmucosal; VFF: von Frey filaments; SMALGO: Small Animal Algometer; CGS: Chronic Gingivo Stomatitis.

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**Table 2. Studies investigating the use of thermal thresholds in experimental cats (in chronological order)**

Study (authors, year and publication type)	Number of cats	Type of algometer	Probe characteristics	Temperature reached (range/ cut off value)	Area tested	Analgesics tested	End-point behavioural response as described by the authors	Baseline thresholds (site and time of measurement)
Casey & Morrow 1983 (research paper)	29	Thermal algometer	Spring-loaded, water-cooled contact thermodes	43-60°C	Shaved outer thighs	NA	Vocalisation, movement of the stimulated limb, interruption of eating/approaching food	NA
Dixon et al. 2002 (research paper)	14	Top Cat Metrology thermal algometer	10 mm long, 10 mm wide and 5 mm deep probe containing a heater element	60°C (cut-off)	Shaved skin of the dorso-lateral thorax	IM meperidine (n=6 out of 14 cats)	Visible (non-defined) reaction of the cat to the application of the stimulus	Shaved thorax before meperidine
Robertson et al. 2003 (research paper)	8	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the thorax	IM saline, morphine, buprenorphine or butorphanol (n=6 cats per group)	Flinching, turning or jumping	Before any treatment
Wegner et al. 2004 (research paper)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the thorax	IV hydromorphone	Flinching, turning or jumping	Before the analgesic treatment
Lascalles & Robertson 2004a (research paper)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the thorax	IV butorphanol (four different doses)	Flinching, turning or jumping	Shaved thorax before the analgesic treatment
Lascalles & Robertson 2004b (research paper)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the thorax	IM hydromorphone, butorphanol and combination of both	Flinching or twitching of the skin, jumping forward, turning to bite the probe	Shaved thorax before any analgesic treatment
Robertson et al. 2005a	6	Top Cat Metrology	As described by Dixon et al.	55°C (cut-off)	Shaved area of	IV and OTM buprenorphine	Flinching, turning or	Shaved thorax

(research paper)		thermal algometer	2002		the thorax	ne	jumping	before any analgesic treatment
Robertson et al. 2005b (research paper)	10	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IV (n=10), TC (n=4), PO (n=2) and IN (n=2) fentanyl	Flinching, turning or jumping	Shaved thorax before any analgesic treatment
Pypendop et al. 2006 (short communication)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IV lidocaine (saline as negative control group)	Jumping, flinching, turning towards the probe, licking or biting the probe area	Shaved thorax before any treatment
Johnson et al. 2007 (research paper)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the thorax	IM butorphanol, buprenorphine and combination of both	Turning to bite the probe, jumping away from the probe, jumping up from a recumbent position	Shaved thorax before any analgesic treatment
Wegner & Robertson 2007 (research paper)	7	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved skin of the lateral thorax	IV hydromorphone	Jumping, flinching or turning toward the probe	Shaved thorax before any analgesic treatment
Slingsby & Taylor 2008 (research paper)	12	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Various shaved areas of the thorax	Buprenorphine (n=12, dexmedetomidine at four different doses (n=10 each) and control saline (n=12)	Skin twitch, jumping or turning head towards the stimulus	Shaved thorax before any treatment
Pypendop et al. 2009 (research paper)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	PO tramadol (versus placebo)	Jumping, turning the head toward the probe, licking or biting the probe area or cable	Shaved thorax before the analgesic treatment
Robertson et al. 2009 (research paper)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	SC hydromorphone	Flinching, jumping or turning to look at the probe	Shaved thorax before the analgesic treatment

Slingsby et al. 2009 (research paper)	12	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IM and OTM dexmedetomidine	Skin twitch, jumping or turning head towards the stimulus	Shaved thorax before the analgesic treatment
Pypendop et al. 2010 (research paper)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	PO gabapentin (versus placebo)	Jumping, turning the head toward the probe, licking or biting the probe area or cable	Shaved thorax before the analgesic treatment
Slingsby et al. 2010 (research paper)	12	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IM buprenorphine (2 different doses), dexmedetomidine (two different doses) and their association (the lowest dose of each)	Skin twitch, jumping or turning the head towards the stimulus	Shaved thorax before drugs administration
Siao et al. 2012 (research paper)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IV oxymorphone and amantadine (oxymorphone and saline as control group)	Jumping, turning the head towards the probe, licking or biting the probe or cable	Shaved thorax before any treatment
Steagall et al. 2013 (research paper)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IV, IM and SC buprenorphine	Flinching, jumping or turning to look at the probe	Shaved lateral thorax before analgesic treatment
Farnworth et al. 2013 a (research paper)	16	Remote carbon dioxide laser	5 mm diameter carbon dioxide beam guided by visible helium laser	Power output 165 mW for all cats	Two shaved areas of the lateral thorax, 4 cm <sup>2</sup> each	NA (validation study)	Moving away from the stimulus or exhibition of the panniculus reflex	NA
Farnworth et al. 2013b (research paper)	113	Remote laser device (Model 48-1, Synrad, Mulkiteo, USA)	3.5 mm diameter carbon dioxide beam guided by visible helium laser	500 mW was used; maximum power output of the device	Two shaved areas of the lateral thorax, 4	NA	Significant shifting (i.e. rising to its feet) or panniculus reflex	NA

set at 10 W cm<sup>2</sup> each

Ambros 2015 (research paper)		Top Cat Metrology thermal algometer (wireless)	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IV hydromorph one or buprenorphi ne followed by a bolus of IV fentanyl (saline as negative control group)	Jumping, flinching, turning towards the probe, licking or biting the probe area	Shaved thorax before any treatment
Farnworth et al. 2015 (research paper)	60	Remote carbon dioxide laser	5 mm diameter carbon dioxide beam guided by visible helium laser	Maximum power output 10 W (cut-off)	Skin of both sides of the thorax	IM morphine, buprenorphi ne, medetomidin e, tramadol, ketoprofen and saline (control)	Significant shifting (i.e. rising to its feet) or panniculus reflex	Skin of both sides of thorax before any treatment
Steagall et al. 2015 (short communication)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	PO codeine and OTM buprenorphi ne	Jumping, flinching, vocalisation, turning towards the probe	Shaved thorax before any treatment
Simon et al. 2016 (research paper)	8	Top Cat Metrology thermal algometer (wireless)	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IV hydromorph one, alone and combined with either butorphanol or buprenorphi ne (saline as negative control group)	Flinching, vocalisation, rolling, jumping, turning the head towards the probe	Shaved thorax before any treatment
Pypendop et al. 2016	8	Top Cat Metrology thermal algometer (wireless)	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	Morphine, methadone or oxymorphon e, administered either IV or OTM	Jumping, turning the head towards the probe, licking or biting the probe or cable	Shaved thorax before any treatment
Taylor et al. 2016 (research paper)	12	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the thorax	SC buprenorphi ne (at three different doses and different formulation)	Skin flick, jumping forward, turning to bite the band, vocalisation	Shaved thorax before any treatment

Doodnaught et al. 2017	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the thorax	PO tapentadol (two different doses) (versus IM buprenorphine as positive control and placebo as negative control)	Vocalisation, rolling, jumping	Shaved thorax after 30 min acclimatisation, before the analgesic treatment
Doodnaught et al. 2018 (Letter to the Editor)	6	Top Cat Metrology thermal algometer	As described by Dixon et al. 2002	Non-specified	Non-specified	OTM buprenorphine	Non-specified	Before the analgesic treatment
Carrozzo et al. 2018 (research paper)	6	Top Cat Metrology thermal algometer (wireless)	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IV fentanyl at two rates of infusion (5 and 3 $\mu\text{g kg hour}^{-1}$ )	Flinching, jumping, turning the head towards the probe, licking or biting the probe area, changing body position	Shaved thorax before the analgesic treatment
Scallan et al. 2019 (research paper)	8	Top Cat Metrology thermal algometer (wireless)	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	Dexmedetomidine either IM or at acupuncture point GV1 (same dose)	Flinching, skin twitch, further dilation of pupils, acute changes in facial conformation, intentional look or motion toward the probe, vocalisation	Shaved thorax before the analgesic treatment
Simon et al. 2019 (research paper)	8	Top Cat Metrology thermal algometer (wireless)	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IV hydromorphone, alone and combined with either butorphanol or naloxone (saline as negative control group)	Flinching, vocalisation, rolling, jumping, turning the head towards the probe	Shaved thorax before any treatment
Simon et al. 2019b (research paper)	10 (all treated at 6,9 and 12 months of age)	Top Cat Metrology thermal algometer (wireless)	As described by Dixon et al. 2002	55°C (cut-off)	Shaved area of the lateral thorax	IV hydromorphone (saline as negative control group)	Flinching, vocalisation, rolling, jumping, turning the head towards the probe	Shaved thorax before any treatment

Table legend: NA: not applicable; GV1: Governing Vessel 1; IV: intravenous; IM: intramuscular; SC: subcutaneous; TT: thermal thresholds; SL: sublingual; TC: transcutaneous (compounded in pluronic lecithin organogel PLO); OTM: oral transmucosal; PO: oral administration; IN: intranasal.

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1 Table 3. Studies investigating (and comparing) different thresholds in experimental cats (in chronological order)

Study (authors, year and publication type)	Number of cats	Type of algometer	Sensitive probe characteristics	Pressure/force /Temperature reached (range/ cut off value)	Area tested	Analgesics tested	End-point behavioural response as described by the authors	Baseline thresholds (site and time of measurement)	Application mode	Main results
Steagall et al. 2006	8	Top Cat Metrology thermal algometer	10 mm long, 10 mm wide and 5 mm deep probe containing a heater element	55 °C (cut-off)	Shaved area of the thorax	SC buprenorphine, morphine, methadone or saline as negative control	Skin flicking, jumping forward, turning to bite the band, vocalisation	Shaved thorax, prior to any treatment administration	Manual, 0.6°C sec <sup>-1</sup> (until end-point response or cut-off reached)	Morphine was the most effective treatment in increasing both TT and MT, as compared to both control group and baseline thresholds
		ProD Plus pressure algometer	Three pins, each tipped with a 2.4 mm diameter ball – bearing pin	650 mmHg (cut-off)	One forearm		Leg shake, head turn and/or vocalisation	Same forearm, prior to any treatment administration	Manual, no time limit (until end-point)	
Steagall et al. 2007 (research paper)	8	Top Cat Metrology thermal algometer	10 mm long, 10 mm wide and 5 mm deep probe containing a heater element	55 °C (cut-off)	Shaved area of the thorax	SC buprenorphine, carprofen or saline as negative control	Skin flicking, jumping forward, turning to bite the band, vocalisation	Shaved thorax, prior to any treatment administration	Manual, 0.6°C sec <sup>-1</sup> (until end-point response or cut-off reached)	Both nociceptive models were effective in detecting buprenorphine analgesia (although the thermal model was superior to the mechanical one), but failed to detect carprofen analgesia
		ProD Plus pressure	Three pins, each tipped	650 mmHg (cut-off)	Craniolateral		Leg shake, head turn	Antebrachium, prior	Manual, no time limit	

		algometer	with a 2.4 mm diameter ball – bearing pin		surface of one antebrahium		and/or vocalisation	to any treatment administration	(until end-point	
Steagall et al. 2008 (research paper)	8 (crossover)	Top Cat Metrology thermal algometer	10 mm long, 10 mm wide and 5 mm deep probe containing a heater element	55 °C (cut-off)	Shaved area of the thorax	SC tramadol, ACP, and their combination (saline as negative control)	Skin flicking, jumping forward, turning to bite the cable	Shaved thorax, prior to any treatment administration	Manual, 0.6°C sec <sup>-1</sup> (until end-point response or cut-off reached)	SC tramadol had limited effect on both TT and MT, whereas the combination ACP + tramadol increased both TT and MT
		ProD Plus pressure algometer	Three pins, each tipped with a 2.4 mm diameter ball – bearing pin	650 mmHg (cut-off)	One forearm		Leg shake, head turn, biting at the probe, vocalisation	Same forearm, prior to any treatment administration	Manual, no time limit (until end-point	
Millette et al. 2008 (short communication)	8	Top Cat Metrology thermal algometer	10 mm long, 10 mm wide and 5 mm deep probe containing a heater element	55 °C (cut-off)	Shaved area of the thorax	IM meperidine (saline as negative control)	Skin flicking, jumping forward, turning to bite the band, vocalisation	Shaved thorax, prior to any treatment administration	Manual, 0.6°C sec <sup>-1</sup> (until end-point response or cut-off reached)	Electrical nociception failed to detect meperidine analgesia, whereas both TT and MT were found useful for this purpose.
		ProD Plus pressure algometer	Three pins, each tipped with a 2.4 mm diameter ball – bearing pin	850 mmHg (cut-off)	Shaved thoracic limb		Picking up and shaking the leg, turning the head towards the bracelet, licking or biting the bracelet, vocalisation	Shaved thoracic limb, prior to any treatment administration	Manual, no time limit (until end-point	
		(Model	Constant Unit	5 mA (cut-	Clipped		Attempts to	Clipped	Continuous	

		CCU1 Constant Current Unit; Astro-Med Inc	Generator equipped with a Grass stimulator, delivering stimuli via two skin electrodes	off)	area of the mid- thorax		look at, lick or bite the electrodes	area of the mid- thorax, prior to any treatment administraction	deliver of stimuli at a rate of 1 mA sec <sup>-1</sup> (duration of 1 ms with 1 ms delay between pulses)	
Ambros et al. 2009 (short communication)	7	Top Cat Metrology thermal algometer	10 mm long, 10 mm wide and 5 mm deep probe containing a heater element	As described by Dixon et al. 2002	As described by Dixon et al. 2002	Epidural hydromorph one (epidural saline as negative control)	As described by Dixon et al. 2002	Pre- treatment thresholds measured	As described by Dixon et al. 2002	Epidural administration of hydromorphone increased both MT and TT values (compared to both saline and baseline values)
		ProD Plus pressure algometer	Three pins, each tipped with a 2.4 mm diameter ball – bearing pin	As described by Dixon et al. 2007	As described by Dixon et al. 2007		As described by Dixon et al. 2007	Pre- treatment thresholds measured	As described by Dixon et al. 2007	
Slingsby et al. 2012 (short communication)	12	Top Cat Metrology thermal algometer	10 mm long, 10 mm wide and 5 mm deep probe containing a heater element	As described by Dixon et al. 2002	As described by Dixon et al. 2002	IM buprenorphi ne, naloxone and their combination	As described by Dixon et al. 2002	Pre- treatment thresholds measured	As described by Dixon et al. 2002	MT were not affected by buprenorphine treatment, whereas TT increased after buprenorphine administration compared to baseline; naloxone antagonised the thermal antinociceptive effect of buprenorphine
		ProD Plus pressure algometer	Three pins, each tipped with a 2.4 mm	As described by Dixon et	As described by Dixon		As described by Dixon et al. 2007	Pre- treatment thresholds	As described by Dixon et al. 2007	

			diameter ball – bearing pin	al. 2007	et al. 2007			measured		
Ambros & Duke 2013 (research paper)	8	Top Cat Metrology thermal algometer	10 mm long, 10 mm wide and 5 mm deep probe containing a heater element	55 °C (cut-off)	Shaved area of the thorax	IV ketamine CRI, delivered for two hours after loading dose, at two different rates	Jumping, flinching, turning towards the probe or licking/biting the probe area	Shaved thorax prior to any treatment administration	Manual, 0.6°C sec <sup>-1</sup> (until end-point response or cut-off reached)	Only the low dose of ketamine minimally affected both TT and MT. The results were inconclusive and ketamine analgesia could not be demonstrated with these nociceptive models
		ProD Plus pressure algometer	Three pins, each tipped with a 2.4 mm diameter ball – bearing pin	20 N (cut-off)	Anterolateral aspect of the antebrachium		Withdrawing, raising or shaking the limb, jumping forwards/turned forwards, trying to bite the actuator	Same forearm prior to any treatment administration	Manual, applying force increasing at 0.8 N sec <sup>-1</sup>	
Addison & Clements 2017 (research paper)	21 (n=14 healthy cats and n=7 cats with OA)	Temperature-controlled thermal aluminium platform	NA	7°C (cold plate) and 40°C (hot plate)	Paws	NA (comparison between healthy cats and cats with OA)	Walking off the plate and number of times and duration that each paw was lifted off the plate	NA	Behavioural observation after 10 second habituation period	MT, measured with both EVF and VFF, were lower in cats with OA than in the healthy ones. Regarding TT, only the cold ones allowed differentiation between healthy and diseased limbs
		EVF and VFF	Probe equipped with 0.8 mm diameter rigid tip (EVF)	Up to 400 g (cut-off EVF) and 0.008-300 g (VFF)	Plantar or palmar aspects of the metacarpal or		Paw withdrawal (prior to filament buckling for VFF)		Manual, no time limit (until end-point response)	

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3 Table legend: SC: subcutaneous; IM: intramuscular; TT: thermal thresholds; MT: mechanical thresholds; ACP: Acepromazine; OA:

4 Osteoarthritis; EVF: Electronic von Frey Anaesthesiometer; VFF: Von Frey Filaments

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