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1	Treatment of Dog	s with Compensated	Myxomatous Mitral	Valve Disease with
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- 2 Spironolactone a Pilot Study
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- 13 Short title: Spironolactone in compensated MMVD
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- 16
- 17 An abstract based on some of these data was presented at the European College of Veterinary
- 18 Internal Medicine (ECVIM) Congress 2012.

19

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31 Treatment of Dogs with Compensated Myxomatous Mitral Valve Disease with

32 Spironolactone – a Pilot Study

33

34 Abstract

Objectives: Spironolactone improves outcome in dogs with advanced myxomatous mitral
valve disease (MMVD). Its efficacy in preclinical MMVD is unknown. Hypothesis;
administration of spironolactone to dogs with compensated MMVD demonstrating risk
factors for poorer prognosis will decrease the rate of disease progression. Aim; to provide
pilot data to evaluate preliminary effects and sample size calculation for a definitive clinical
trial.

Animals: Twenty-five client-owned dogs with MMVD with at least one of the following; left
atrial-to-aortic ratio (LA:Ao) ≥1.5, normalized left ventricular internal diameter in diastole
(LVIDdN) ≥1.6), N-terminal pro-B-type natriuretic peptide (NT-proBNP) >550pmol/L,
cardiac troponin I (cTnI) >0.025ng/mL.

45 Methods: Prospective, single-center, equally randomized, placebo-controlled, double-blinded,
46 parallel grouped pilot study. No dogs were receiving medications for cardiac disease prior to
47 enrolment.

Results: Twelve dogs received placebo; 13 received spironolactone. One dog in the spironolactone group died suddenly, 1 developed congestive heart failure and 2 received suboptimal spironolactone doses. At enrolment NT-proBNP was significantly higher in the spironolactone group (P=0.005). LA:Ao (P=0.002) and LVIDdN (P=0.005) increased over time in the placebo group, but not the spironolactone group; the change did not differ significantly between groups. The change in biomarker concentrations did not differ significantly between groups; there was a tendency towards an increase in NT-proBNP over

- time in the placebo group. Enrolment of 76 dogs would be necessary to demonstrate a
- 56 difference in the change in LA:Ao over 6 months between groups.
- 57 **Conclusions:** preliminary results support undertaking a larger clinical trial of treatment of
- 58 dogs with preclinical MMVD with spironolactone.
- 59

60 Keywords: preclinical disease, therapy, canine

- 61
- 62 Abbreviations:

ACVIM	American College of Veterinary Internal Medicine
Ао	Aorta
CHF	Congestive heart failure
CKCS	Cavalier King Charles spaniel
cTnI	Cardiac troponin I
ECG	Electrocardiogram
ELISA	Enzyme-linked immunosorbent assay
LA	Left atrium
LA:Ao	Ratio of left atrial to aortic root diameter
LVIDd	Left ventricular internal dimension in diastole
	Ratio of left ventricular end-diastolic dimension to left
LVIDd/ LVFWd	ventricular free wall thickness in diastole

	Left ventricular end-diastolic dimension normalized for body
LVIDdN	weight
LVIDs	Left ventricular internal dimension in systole
	Left ventricular end-systolic dimension normalized for body
LVIDsN	weight
LVFWd	Left ventricular free wall thickness in diastole
MMVD	Myxomatous mitral valve disease
NSAID	Non-steroidal anti-inflammatory drug
NT-proBNP	N-terminal pro-B-type natriuretic peptide
PCV	Packed cell volume
UAC	Urinary aldosterone to creatinine ratio

65 Myxomatous mitral valve disease (MMVD) is the commonest cause of cardiovascular 66 disease in the dog [1, 2]. Valvular degeneration results in regurgitation of blood, leading to progressive volume overload of the left atrium and ventricle, which compensate by eccentric 67 68 hypertrophy [3]. The American College of Veterinary Internal Medicine (ACVIM) classification system for MMVD describes four disease stages, from A (at risk) to D 69 70 (decompensated congestive heart failure) [4]. The compensated, preclinical stage of the 71 disease (stage B) is subdivided according to the absence (stage B1) or presence (stage B2) of 72 evidence of compensatory hypertrophy, identified on the basis of chamber enlargement on radiography or echocardiography. The rate of progression of the disease is variable and the 73 74 development of heart failure is not an inevitable consequence of MMVD [5]. However, even with optimal medical therapy, once dogs with MMVD develop congestive heart failure 75 76 (CHF), median survival is approximately 270 days [6]. Pimobendan has been shown to delay the onset of congestive heart failure in dogs with ACVIM class B2 MMVD [7]. Further 77 78 therapeutic strategies, targeting different mechanisms of disease progression, might provide 79 additional clinical benefit in dogs at risk of disease progression.

80 The identification of dogs at greatest risk of progression of MMVD is important in a disease with such variability of outcome. Evidence of cardiac remodelling, such as increased 81 echocardiographic measurements of left atrial [5] and left ventricular size, [8] are associated 82 83 with decreased survival times in dogs with MMVD. Increases in secondary markers of myocardial stress and injury, specifically, serum N-terminal pro-B-type natriuretic peptide 84 85 $(NT-proBNP) > 524 \text{ pmol/L and/ or serum cardiac troponin I (cTnI) > 0.025 ng/mL, are also$ associated with poorer outcomes [9]. Increased left atrial size is associated with an increased 86 risk of disease progression for dogs in ACVIM class B [10]. 87

88 Activation of the renin-angiotensin-aldosterone system is important in the pathophysiology

89 of cardiac remodelling in canine MMVD [11]. Via its actions on the mineralocorticoid receptor, aldosterone promotes fluid retention, leading to volume-overload and stimulates 90 91 myocardial fibrosis [12]. Urinary aldosterone to creatinine ratio (UAC) is associated with the 92 rate of change of left ventricular size in dogs with MMVD, suggesting that aldosterone production increases during periods of active remodelling [13]. Spironolactone is a 93 94 mineralocorticoid receptor antagonist that has been shown to prolong survival times in dogs with advanced MMVD and CHF secondary to MMVD, when given in combination with 95 96 standard therapy [14]. The use of spironolactone in dogs with compensated MMVD has not 97 been reported, although a study investigating its effects in combination with benazepril is currently ongoing. 98

We hypothesized that chronic oral administration of spironolactone to dogs with compensated MMVD demonstrating risk factors known to be associated with decreased survival times (increased left atrial and ventricular size and increased serum NT-proBNP and cTnI), not receiving any other cardiovascular medications, would result in decreased rates of change of these risk factors over time. The aim of the study was to provide pilot data to evaluate preliminary effects, drug safety and to calculate the number of dogs needed for a definitive clinical trial.

106

107 Animals, Materials and Methods

108 *Study Design*

109 The design of this pilot study was single-centre, prospective, equally randomized, double-110 blinded, parallel-grouped and placebo-controlled. Each dog participated in the study for a 111 period of six months. The study was conducted in the United Kingdom. The study was 112 approved by the Royal Veterinary College Ethical Committee and specific informed owner 113 consent was obtained (unique reference number 2010 1039).

114

115 *Dogs*

116 Client-owned dogs of a variety of breeds with echocardiographically-confirmed MMVD were 117 prospectively recruited between December 2010 and December 2013 from those already 118 enrolled in a longitudinal study of canine MMVD in first opinion practice [8]. Dogs were referred to the longitudinal study by the veterinarians at 2 London-based first opinion 119 120 practices after detection of a murmur consistent with mitral regurgitation at any stage in the 121 natural history of the disease. Echocardiography was performed to confirm the diagnosis of 122 MMVD and to exclude the presence of other cardiac diseases. Diagnosis of MMVD was on 123 the basis of characteristic abnormalities of the valve leaflets (thickening, prolapse, or both) and evidence of regurgitant flow across the valve detected by Doppler. Dogs with any other 124 125 cardiac disease or clinically relevant organ-related or systemic disease were not enrolled in the longitudinal study. 126

127 To be eligible for inclusion in the present study, a dog had to have echocardiographic evidence of MMVD, defined as above, and demonstrate at least one risk factor for disease 128 129 progression (evidence of cardiomegaly (defined as left atrial to a rtic ratio (LA:Ao) ≥ 1.5 and/ or left ventricular end-diastolic dimension, normalized for body weight (LVIDdN)) > 130 1.6 [8], serum NT-proBNP > 550pmol/L [9] and/ or serum cTnI > 0.025 ng/mL [9]. During 131 132 the screening process, but not during the experimental phase, serum NT-proBNP was measured using the first-generation version of a commercially-available enzyme-linked 133 134 immunosorbent assay (ELISA).^c

Dogs were excluded from the study if they had any of the following: evidence of anycongenital or acquired cardiac disease other than MMVD; evidence of kidney disease,

hypoadrenocorticism (on the basis of historical, physical examination and routine
biochemical findings; ACTH stimulation tests were not performed), hyperkalemia, or
hypernatremia; current or previous clinical signs of congestive heart failure or current
medical therapy for cardiac disease. The summary of product characteristics for Prilactone
recommends that dogs treated concomitantly with spironolactone and non-steroidal antiinflammatory drugs (NSAIDs) be correctly hydrated,^d and so it was recommended that dogs
should not receive NSAIDs, although this was not an absolute exclusion criterion.

144

145 *Randomization*

146 Randomization was by patient (dog). The study was initially designed to recruit 20 dogs. Prior to the enrolment phase, a numbered list of 20 random group assignments (either group 147 A or group B) was compiled by drawing assignments from a hat. Dogs were assigned to 148 149 groups according to the order in which they were enrolled in the study. Prospectively determined, prognostic factor balance was achieved by minimization to ensure that the 150 151 number of cavalier King Charles spaniels (CKCS) was equal in each group [15]; briefly, if, in the latter stages of the recruitment phase (from dog number 11 onwards), enrolling a CKCS 152 to the next group assignment according to the randomization list would have resulted in 153 unbalancing of the groups, then the dog was assigned to the alternative treatment group. No 154 other variable was considered prior to enrolment. Data obtained from the first 20 dogs were 155 analysed and reported in abstract form^e and the decision made to recruit an additional 20 dogs 156 157 to increase the statistical power of the study. An additional numbered list of 20 random 158 assignments (either group C or group D) was compiled in the same way to allow recruitment of additional dogs to the study in a blinded fashion after treatment allocation of groups A and 159 160 B was revealed at the time of previous data analysis.

162 Blinding

The investigators and owners were blinded to the treatment allocation. Assignment of enrolled dogs to treatment groups was performed by a veterinary nurse to conceal allocation from the investigators responsible for measurement of the variables of interest. Data for the first 20 dogs enrolled (groups A and B) was analyzed prior to recruitment of the additional dogs (groups C and D).^e The blinding codes for the treatment groups were held by the sponsor until the time of each data analysis.

169

170 Trial medication

Spironolactone verum (Prilactone 10 mg tablets)^f was administered orally at a target dose of 2 171 172 mg/kg SID, as per registered label instructions, and the dose adjusted to a suitable number of tablets. Placebo was administered PO according to the calculated daily dose for 173 spironolactone verum tablets and adjusted to a suitable number of placebo tablets. The 174 175 tablets and packaging of the verum and placebo were visually indistinguishable. Dogs received tablets (verum or placebo) orally once daily for 6 months, unless otherwise stated. 176 The dose of the study medication was not adjusted during the study period. Additional 177 178 appropriate medication was prescribed if clinical signs of cardiac failure developed during the study period. Participation was terminated if clinical signs of another significant medical 179 condition occurred which required additional treatment or warranted euthanasia, or if adverse 180 181 effects were observed which necessitated cessation of the therapy. A record was kept of other 182 medication used. Compliance was monitored by counting the number of unused pills returned 183 by the owner at the end of the study period. This number was compared with the expected number of pills remaining. 184

186 Schedule of Events

187 Prior to enrolment, serum biochemistry and electrolyte measurements were performed. 188 At baseline, enrolled cases underwent a full evaluation, comprising recording of the history, 189 measurement of systolic arterial blood pressure by Doppler sphygmomanometry,^g physical 190 examination, blood sampling, electrocardiography (ECG) and echocardiography, in that order. Electrocardiography was performed in right lateral recumbency and heart rate was 191 192 measured from a 60-second recording of lead II. Treatment was initiated with either verum or 193 placebo. Re-examinations were scheduled at day 14 and approximately 6 months after 194 inclusion. The tests performed at each study visit are summarized in Figure 1. 195 196 Clinical Evaluation 197 At inclusion, demographic characteristics (age, breed, sex and neutering status) were 198 recorded. Body weight and body condition score were recorded at each study visit. 199

200 Blood sampling and Laboratory Analysis

Blood was collected by jugular venepuncture into serum gel tubes and K3-EDTA-treated
tubes. Free-catch urine samples were collected. Samples were chilled at 4°C for up to 6 hours
before separation by centrifugation. Packed cell volume (PCV) was measured prior to
separation. Serum and urine samples were transported to a commercial laboratory for
measurement of routine biochemical parameters and electrolytes.^h The remaining serum and
urine were stored at -80°C for batched analysis. Urinary aldosterone concentrations were
measured using a previously-validated, commercially-available radioimmunoassay [16]

208 following mild acid hydrolysis and extraction into ethyl acetate, as previously described [13]. 209 Serum aliquots were transported to the same commercial laboratory on dry ice. Before 210 analysis, the frozen serum was allowed to thaw slowly at room temperature. Concentrations of cTnI were measured using an ELISAⁱ according to the manufacturer's instructions. The 211 use of this assay has been previously validated for canine samples [17]. Concentrations of 212 213 NT-proBNP were measured using the second-generation version of a previously-validated canine NT-proBNP ELISA^c according to the manufacturer's instructions [18]. Serum 214 biochemistry and electrolytes, NT-proBNP, cTnI, PCV and UAC were measured at the 215 216 baseline visit. On day 14, PCV, serum biochemistry and electrolytes and UAC were 217 measured. Full clinical evaluation, plus measurement of PCV, serum biochemistry and 218 electrolytes, NT-proBNP, cTnI and UAC, was repeated at the 6 month time point, unless 219 otherwise stated.

220

221 Echocardiography

222 Echocardiography was performed at baseline and at the 6 month visit. Echocardiographic examinations were performed by a single board-certified cardiologist (AB). Dogs were placed 223 in right and then left lateral recumbency on an ultrasound examination table. The 224 225 echocardiographic examination was performed using an ultra-sound unit^j equipped with 2-4 226 MHz and 3–7 MHz phased array transducers and ECG monitoring. Standard imaging planes 227 were digitally stored. Assessment of mitral valve structures was performed from the right 228 parasternal long-axis view and the left apical 4-chamber view. The LA:Ao was measured 229 from the right parasternal short axis view, as previously described [19]. Left ventricular internal diameters in systole and diastole (LVIDs and LVIDd, respectively) and wall 230 231 thicknesses were measured from M-mode obtained from the right parasternal short axis view. 232 LVIDs was normalized for body weight (LVIDsN) by the formula: LVIDs/ (body weight

[kg])^{0.315}[20]. LVIDd was normalized for body weight (LVIDdN) by the formula: LVIDd/
(body weight [kg])^{0.294}[20]. The ratio of LVIDd to left ventricular free wall thickness in
diastole (LVFWd) was calculated (LVIDd/ LVFWd) as an indirect estimate of wall stress.
Measurements were recorded from at least 3 cardiac cycles and the mean value used in
subsequent analyses.

The primary outcome comparisons of the study were comparisons of the change in LA:Ao, 238 LVIDdN and serum NT-proBNP and cTnI between groups over a 6 month period. Secondary 239 240 outcome comparisons were comparisons of the change in other variables (PCV, serum urea, creatinine and electrolyte concentrations, UAC, LVIDsN, LVIDd/ LVFWd ratio, E wave 241 velocity, E/A wave ratio, heart rate and body weight) between groups over the same 6 month 242 243 period and between-group comparisons at different time points (baseline and 2 and 26 weeks after enrolment). Results are reported according to the Consolidated Standards of Reporting 244 245 Trials 2010 guidelines for reporting parallel group randomized trials [21].

246

247 Statistical Analysis

Data were analyzed on an intention to treat basis and data from all dogs that were randomly 248 249 assigned were included. Statistical analyses were performed using commercially-available software.^k Data were assessed for normality graphically and by use of the Shapiro-Wilk test. 250 251 Results are reported as mean ± standard deviations for normally distributed continuous 252 variables or median [range] for non-normally distributed variables. Repeated measures linear mixed models with the random effect of subject (dog) were constructed to compare the 253 254 change in variables over time, and a compound symmetry (co)variance structure was 255 assumed between residuals of the same subject (dog) in the model. The effects of treatment 256 group (spironolactone vs. placebo), time (treated as a continuous covariate) and interaction

257 between treatment group and time were included in the model. Residuals were assessed 258 graphically for normality. Variables were logarithmically transformed if the residuals were 259 not normally distributed. The assumption of homogeneity of variance was tested by plotting 260 the predicted values against the residual values. Comparisons of continuous variables between groups at different time points were made using independent t-tests or Mann-261 262 Whitney U tests, as appropriate. Fisher's exact tests were used to compare proportions between groups at baseline. A value of P ≤ 0.05 was considered significant. In view of the 263 small sample size and the pilot nature of the study a value of P < 0.1 was considered to 264 265 indicate a tendency towards significance. A sample size calculation was performed on the basis of the data obtained from the present pilot study using commercially-available software, 266 267 assuming $\alpha = 0.05$ and $\beta = 0.2$ (i.e. power = 0.8).¹

268

269 <u>Results</u>

270 Progress through the phases of the study is summarized in Figure 2. Twenty-five dogs with 271 compensated MMVD diagnosed on the basis of echocardiographic findings were enrolled in the study. Due to the low rate of suitable dogs presenting to the larger longitudinal study, it 272 proved impossible to recruit 40 dogs. Twenty-four dogs had LVIDdN > 1.6 and/ or LA:Ao > 273 274 1.5. Fifteen dogs had a previous measurement of serum cTnI > 0.025 pmol/L. Twenty-one 275 dogs had a previous measurement of serum NT-proBNP > 550 pmol/L. All dogs met the 276 inclusion criteria; 13 dogs demonstrated all three risk factors, 8 dogs demonstrated two of the 277 risk factors and 4 dogs demonstrated one risk factor. Baseline characteristics of the two 278 groups are presented in Table 1. There was no evidence for differences in age, body weight, gender or proportions of CKCS to other breeds between groups. Serum NT-proBNP 279 280 concentrations were higher in the spironolactone treatment group compared with the placebo 281 group (P = 0.005). At baseline there was a tendency for serum potassium (P = 0.078) to be

higher in the spironolactone treatment group compared with the placebo group. Group-wisecomparisons were otherwise unremarkable. No evidence of kidney disease,

hypoadrenocorticism, hyperkalemia, or hypernatremia was detected in any dog on serumbiochemical and electrolyte analysis.

Twelve dogs (5 neutered females, 2 entire males and 5 neutered males) with ages ranging 286 from 6.3 to 13.1 years and body weights ranging from 4.5 to 18.2 kg were assigned to receive 287 288 placebo. These dogs comprised 7 CKCS, 2 mixed breeds and 1 each of bichon frisé, lurcher 289 and toy poodle. No compliance problems, potential adverse drug reactions or adverse events 290 were reported for dogs in this group. Two dogs had been receiving NSAIDs prior to, but not 291 at the time of, recruitment. NSAID therapy was reinitiated by the primary veterinarian during 292 the trial period in one of these dogs. NSAID therapy was initiated during the trial period in a third dog. 293

294 Thirteen dogs (2 neutered females and 11 neutered males) with ages ranging from 6.1 to 295 13.4 years and body weights ranging from 1.8 to 23.3 kg were assigned to receive 296 spironolactone. These dogs comprised 8 CKCS, 2 cross-breeds and 1 each of bichon frisé, collie and Chihuahua. One dog in this group received a suboptimal dose of trial medication 297 (1.3 mg/kg once daily) throughout the trial period, due to owner non- compliance. One dog 298 299 was treated for seasonal allergic dermatitis during the trial period, during which time the trial 300 medication was withdrawn for 10.6 weeks. Trial medication was reinstated following 301 resolution of the dermatitis, which did not subsequently recur.

Two dogs in the spironolactone group suffered adverse events during the trial period (one dog developed congestive heart failure requiring medical management and one dog died suddenly). The dog that died suddenly was not known to be in congestive heart failure prior to death. There was no difference in the proportion of dogs experiencing adverse events between groups (P = 0.480). One dog had been receiving NSAIDs prior to, but not at the time of, recruitment. NSAID therapy was reinitiated by the primary veterinarian during the
trial period; this was the dog that died suddenly, although the death was considered to be
unrelated to NSAID therapy.

310 Results of repeated measures linear model analyses for the primary outcome comparisons 311 are summarized in Table 2. Residuals were not normally distributed for serum cTnI. Following logarithmic transformation of this variable the residuals were normally distributed. 312 313 Because baseline serum NT-proBNP measurements were higher in the group receiving 314 spironolactone, baseline measurements were included in the model as a covariate for this 315 variable. No significant differences were detected for the change over time of any variable 316 between groups. The change in serum NT-proBNP tended to be greater for the placebo group 317 (P = 0.087). There was a tendency for serum NT-proBNP concentrations to increase in the placebo group, but not in dogs receiving spironolactone (P = 0.073) (Figure 3). Left atrial to 318 319 aortic ratio (P = 0.002) and LVIDdN (P = 0.005) increased over time in the placebo group, 320 but not in dogs receiving spironolactone ((P = 0.231 and P = 0.194, respectively, Figures 4)321 and 5); however, in the absence of significant differences in the change over time between 322 groups these findings should be interpreted with caution.

323 Results of repeated measures linear model analyses for the secondary outcome comparisons 324 are summarized in Table 3. Residuals were not normally distributed for UAC, serum 325 creatinine or E wave velocity. Following logarithmic transformation of these variables the 326 residuals were normally distributed. There was a tendency for LVIDd/ LVFWd ratio to increase over time in the placebo group (P = 0.070), but not in dogs receiving spironolactone 327 (P = 0.315). The change in LVIDd/ LVFWd ratio over time was not different between 328 329 groups. There was a tendency for body weight to decrease in the dogs receiving spironolactone (P = 0.066), but not in the placebo group. The change in body weight was not 330 331 different between groups.

Comparisons between groups at the 2 week time point are summarized in Table 4. No dogs were withdrawn from the study at this time point due to pharmacovigilance concerns. Urinary aldosterone to creatinine ratio was significantly higher in dogs receiving spironolactone than those receiving placebo (P = 0.006). There was a tendency for serum potassium to be higher in dogs receiving spironolactone than those receiving placebo (P = 0.098). There was no evidence of differences in any other variable between groups at this time point.

Comparisons between groups at the 6 month time point are summarized in Table 5. There was a tendency for serum NT-proBNP to be higher in dogs receiving spironolactone than those receiving placebo (P = 0.051). There was no evidence of differences in any other variable between groups at this time point.

A post-hoc sample size calculation was performed using the mean change in LA:Ao observed from these preliminary data, which suggested that a total sample of 76 dogs (38 per group) would be necessary to demonstrate a significant difference in the change in LA:Ao over 6 months between groups if one existed. However, use of the 95% confidence intervals for the change in LA:Ao for the sample size calculation suggested that the range of total sample size required to demonstrate a difference is 36 to 2936 dogs.

348

349 <u>Discussion</u>

The results of the present study suggest that a larger sample size (76 dogs) would be necessary to definitively test the hypothesis that spironolactone slows the rate of progression of preclinical MMVD. Although none of the between groups comparisons reached statistical significance, this pilot study is likely to be underpowered to demonstrate such differences. Four dogs receiving spironolactone failed to adhere to the protocol throughout the entire six month study period; one died suddenly, one developed congestive heart failure necessitating 356 the addition of other therapy, one received a suboptimal dose of medication and 357 administration of trial medication was suspended in one dog for part of the trial period. This 358 is likely to have further reduced the power of the study to detect differences between the 359 groups. A post hoc sample size calculation was performed in order to inform the design of a subsequent, larger study that could be performed to determine whether a real treatment effect 360 361 exists. Given the 95% confidence interval of the average change in LA:Ao, the number of dogs required could be as few as 36 or as many as 2936. Because of the intrinsic unreliability 362 363 of such a sample size prediction on the basis of data from a relatively small population it is 364 possible that there is no treatment effect, in which case no difference between groups would be demonstrated even if an infinite number of dogs was studied. No definitive conclusions 365 366 regarding any effect of spironolactone on disease progression in dogs with MMVD can 367 therefore be drawn on the basis of the results of this pilot study.

Echocardiographic measurements of cardiac size (LA:Ao and LVIDdN) increased over 368 time in the placebo group but not in dogs receiving spironolactone, although no difference in 369 the change over time was detected between groups. This suggests that performing a larger, 370 371 definitive clinical trial is warranted to further investigate whether treatment with spironolactone slows disease progression in dogs with preclinical MMVD. Increasing LA:Ao 372 and LVIDdN are known to be associated with reduced survival times in dogs with MMVD 373 374 [22]. It is possible that decreasing the rate of increase of cardiac size might delay the onset of congestive heart failure, although large, long-term studies are required to investigate this 375 hypothesis. 376

The radiographic indices of cardiac size in CKCS with compensated MMVD increase in a non-linear fashion with disease progression, reaching their maximal rate of change in the 9 months prior to the onset of congestive heart failure [23]. It was expected, therefore, that in this population of dogs with compensated MMVD and risk factors likely to be associated 381 with progression, echocardiographic indices of cardiac size would increase significantly over a six month period. The observation that LA: Ao and LVIDdN increased significantly in the 382 383 placebo group suggests that this expectation was reasonable. Patients with MMVD that have 384 higher NT-proBNP concentrations and greater heart size tend to have more advanced disease. Thus, since the group of dogs receiving spironolactone had significantly higher NT-proBNP 385 386 concentrations at baseline it might have been expected that these dogs would have a more rapid rate of change than those in the placebo group. This might have contributed to the 387 failure of the study to demonstrate a difference in the change over time in LA: Ao and 388 389 LVIDdN. Future studies could use stratification of randomization of recruited cases by echocardiographic indices of heart size and/ or serum NT-proBNP concentrations to ensure 390 391 that treatment groups are balanced with respect to these variables. In a small pilot study this 392 approach was not feasible.

393 Urinary aldosterone to creatinine ratio was higher in dogs receiving spironolactone compared with placebo at the 2 week time point but not at the 6 month time point. 394 Mineralocorticoid receptor blockade by spironolactone increases aldosterone secretion by 395 396 stimulation of renin production [24]. It was expected that UAC would remain higher in the 397 group receiving spironolactone throughout the study period. However, data were missing for this variable on 21/69 (30.4%) occasions on which it should have been measured due to 398 399 failure to obtain urine samples. This would have decreased the power of the study to detect significant differences in UAC between treatment groups. 400

There was a tendency for serum potassium concentrations to be higher in dogs treated with spironolactone at baseline and the 2 week time point. Nevertheless, all measurements remained within the range of normal values and so this is not a source of pharmacovigilance concern nor has it been noted as a clinically significant adverse event in the published clinical trials involving spironolactone in dogs [25]. There was no evidence that the serum cTnI concentrations changed over time in either treatment group. Rapid increases in serum cTnI concentrations only occur late in the course of the disease, within the last 6 months of life of dogs that die due to MMVD [9]. A rapid increase in serum cTnI would not, therefore, be expected in dogs in the compensated phase of the disease.

Fractional shortening is usually increased in the compensated phase of MMVD, as the presence of mitral insufficiency results in altered loading conditions. LVIDsN does not, therefore, increase until the late stages of MMVD [22, 26]. In a similar fashion to serum cTnI, it would not therefore be expected that LVIDsN would rapidly increase over a six month period in this population of dogs.

Increased E wave velocities are indicative of increased ventricular filling pressures and the volume of the regurgitant jet [26]. The results of the present study suggest that ventricular filling pressures were not increasing and ventricular diastolic function was not declining in either treatment group, probably reflecting the relatively early stage of the disease in this population. E wave velocities >1.2 m/s have previously been shown to be associated with an increased risk of disease progression in dogs with ACVIM class B MMVD; [10] the majority of dogs in the present study had E wave velocities <1.2 m/s at the baseline visit.

423 This study has a number of limitations. Firstly, the sample size calculation suggests that on 424 the basis of our own data 76 dogs would need to be studied in order to demonstrate a 425 difference in the change in LA: Ao over 6 months between groups if one exists. This pilot 426 study was therefore underpowered to detect such a difference. We have therefore referred to 427 tendencies towards significance in the data with a more lenient P-value of < 0.1, while 428 recognizing the risks of a type I statistical error inherent in this approach. Secondly, no 429 consensus exists with regard to determining optimal cut-offs for echocardiographic evidence 430 of cardiomegaly in canine MMVD. In the present study, LA:Ao > 1.5 and / or LVIDdN >

431 1.6, were used to select dogs at risk of progression. These represent relatively liberal criteria, as the upper 95% confidence interval for LVIDdN is 1.85 [20]. However, in the present 432 433 study these cut-offs were used only as inclusion criteria, in an attempt to identify dogs with 434 more advanced compensated MMVD that were more likely to experience disease progression over a 6 month period. Previous studies have demonstrated that dogs above cut-off values 435 436 lower than the value of 1.85 are at increased risk of cardiac related mortality [8]. Thirdly, increases in serum cTnI are not specific for cardiac disease, [27, 28] and so use of this marker 437 as an inclusion criterion has limitations. Nevertheless, no dog was recruited solely on the 438 basis of the cTnI measurement. 439

Fourthly, this study included a high proportion of CKCS. Urinary aldosterone to creatinine 440 441 ratio has been shown to be higher in CKCS than non-CKCS breeds [13]. It is possible, therefore, that any benefit of mineralocorticoid receptor blockade might be greater in CKCS 442 than other breeds. For this reason, the groups were balanced for numbers of CKCS by the 443 minimization method; the trial was not, therefore, truly randomized. However, 444 mineralocorticoid receptor antagonists have been shown to be beneficial in human patients, 445 446 regardless of plasma aldosterone concentrations [29]. Large studies that allow for subanalyses according to breed are necessary to investigate whether breed- specific effects of 447 spironolactone exist. 448

Finally, allocation concealment would ideally have been performed externally rather than
by a member of the regular clinic team. Inadequate allocation concealment may lead to
selection bias and hence produce spurious results. In the present study, however, every
attempt was made to maintain allocation concealment and therefore avoid bias of this nature.

453

454 <u>Conclusions</u>

- 455 The preliminary findings of this pilot study support undertaking a larger, definitive,
- 456 prospective, randomized, placebo-controlled, double-blinded clinical trial to further evaluate
- the effect of spironolactone on disease progression in dogs with preclinical MMVD.
- 458
- 459 Footnotes
- 460 ^c Canine Cardiopet Nt-proBNP, IDEXX Laboratories, Westbrook, ME
- 461 ^d http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_-
- 462 _Product_Information/veterinary/000105/WC500063372.pdf
- 463 ^e M.J. Hezzell, A. Boswood, J. Elliott. Treatment of dogs with compensated degenerative
- 464 mitral valve disease (DMVD) with spironolactone. Journal of Veterinary Internal Medicine
- **465** 2012; 26:1517.
- 466 ^f CEVA Santé Animale, Libourne, France
- 467 ^g Park model 811 B, Perimed, Bury St. Edmonds, UK
- 468 ^h IDEXX Laboratories, Wetherby, UK
- 469 ⁱ Access Systems AccuTnI Assay, Beckman Coulter Inc., Fullerton, CA
- 470 ^j Acuson Cypress, Siemens Medical Solutions, Siemens House, Oldbury, Bracknell, UK
- 471 ^k IBM SPSS version 23, SPSS Inc., Chicago, IL
- 472 ¹GLIMMPSE version 2.2.4, University of Colorado, Denver, CO

473	References

475 1. Buchanan J. Prevalence of cardiovascular disorders. In: Fox P, Sisson D, Moise N, editors. Textbook of Canine and Feline Cardiology. 2nd ed. Philadelphia: W.B. Saunders; 476 1999. p. 457-70. 477 478 2. Egenvall A, Bonnett BN, Hedhammar A, Olson P. Mortality in over 350,000 insured Swedish dogs from 1995-2000: II. Breed-specific age and survival patterns and relative risk 479 for causes of death. Acta Vet Scand. 2005;46:121-36. 480 481 3. Kittleson MD, Kienle RD. Myxomatous atrioventricular valvular degeneration. In: Kittleson MD, Kienle RD, editors. Small Animal Cardiovascular Medicine. 1st ed. St. Louis: 482 483 Mosby; 1998. p. 297-318. 4. 484 Atkins C, Bonagura J, Ettinger S, Fox P, Gordon S, Haggstrom J, Hamlin R, Keene B, Luis-Fuentes V, Stepien R. Guidelines for the diagnosis and treatment of canine chronic 485 valvular heart disease. J Vet Intern Med. 2009;23:1142-50. 486 487 5. Borgarelli M, Savarino P, Crosara S, Santilli RA, Chiavegato D, Poggi M, Bellino C, La Rosa G, Zanatta R, Haggstrom J, Tarducci A. Survival characteristics and prognostic 488 489 variables of dogs with mitral regurgitation attributable to myxomatous valve disease. J Vet Intern Med. 2008;22:120-8. 490 491 6. Haggstrom J, Boswood A, O'Grady M, Jons O, Smith S, Swift S, Borgarelli M, 492 Gavaghan B, Kresken JG, Patteson M, Ablad B, Bussadori CM, Glaus T, Kovacevic A, Rapp M, Santilli RA, Tidholm A, Eriksson A, Belanger MC, Deinert M, Little CJ, Kvart C, French 493 494 A, Ronn-Landbo M, Wess G, Eggertsdottir AV, O'Sullivan ML, Schneider M, Lombard CW,

495 Dukes-McEwan J, Willis R, Louvet A, DiFruscia R. Effect of pimobendan or benazepril

496 hydrochloride on survival times in dogs with congestive heart failure caused by naturally

497 occurring myxomatous mitral valve disease: the QUEST study. J Vet Intern Med.

498 2008;22:1124-35.

499 7. Boswood A, Haggstrom J, Gordon SG, Wess G, Stepien RL, Oyama MA, Keene BW, Bonagura J, MacDonald KA, Patteson M, Smith S, Fox PR, Sanderson K, Woolley R, 500 501 Szatmari V, Menaut P, Church WM, O'Sullivan ML, Jaudon JP, Kresken JG, Rush J, Barrett KA, Rosenthal SL, Saunders AB, Ljungvall I, Deinert M, Bomassi E, Estrada AH, Fernandez 502 503 Del Palacio MJ, Moise NS, Abbott JA, Fujii Y, Spier A, Luethy MW, Santilli RA, Uechi M, 504 Tidholm A, Watson P. Effect of Pimobendan in Dogs with Preclinical Myxomatous Mitral 505 Valve Disease and Cardiomegaly: The EPIC Study-A Randomized Clinical Trial. J Vet Intern Med. 2016;30:1765-1779. 506 507 8. Moonarmart W, Boswood A, Luis Fuentes V, Brodbelt D, Souttar K, Elliott J. N-508 terminal pro B-type natriuretic peptide and left ventricular diameter independently predict mortality in dogs with mitral valve disease. J Small Anim Pract. 2010;51:84-96. 509 9. 510 Hezzell MJ, Boswood A, Chang YM, Moonarmart W, Souttar K, Elliott J. The 511 combined prognostic potential of serum high-sensitivity cardiac troponin I and N-terminal pro-B-type natriuretic peptide concentrations in dogs with degenerative mitral valve disease. 512 513 J Vet Intern Med. 2012;26:302-11. 10. Borgarelli M, Crosara S, Lamb K, Savarino P, La Rosa G, Tarducci A, Haggstrom J. 514 Survival characteristics and prognostic variables of dogs with preclinical chronic 515 516 degenerative mitral valve disease attributable to myxomatous degeneration. J Vet Intern Med.

517 2012;26:69-75.

518 11. Pedersen HD, Koch J, Poulsen K, Jensen AL, Flagstad A. Activation of the renin519 angiotensin system in dogs with asymptomatic and mildly symptomatic mitral valvular
520 insufficiency. J Vet Intern Med. 1995;9:328-31.

521 12. Tan LB, Schlosshan D, Barker D. Fiftieth anniversary of aldosterone: from discovery
522 to cardiovascular therapy. Int J Cardiol. 2004;96:321-33.

Hezzell MJ, Boswood A, Chang YM, Moonarmart W, Elliott J. Associations among
serum N-terminal procollagen type III concentration, urinary aldosterone-to-creatinine ratio,
and ventricular remodeling in dogs with myxomatous mitral valve disease. Am J Vet Res.
2012;73:1765-74.

527 14. Bernay F, Bland JM, Haggstrom J, Baduel L, Combes B, Lopez A, Kaltsatos V.

528 Efficacy of spironolactone on survival in dogs with naturally occurring mitral regurgitation

529 caused by myxomatous mitral valve disease. J Vet Intern Med. 2010;24:331-41.

Taves DR. Minimization: a new method of assigning patients to treatment and control
groups. Clin Pharmacol Ther. 1974;15:443-53.

- 532 16. Syme HM, Fletcher MG, Bailey SR, Elliott J. Measurement of aldosterone in feline,
 533 canine and human urine. J Small Anim Pract. 2007;48:202-8.
- 534 17. Oyama MA, Solter PF. Validation of an immunoassay for measurement of canine
 535 cardiac troponin-I. J Vet Cardiol. 2004;6:17-24.
- 536 18. Fox PR, Oyama MA, Hezzell MJ, Rush JE, Nguyenba TP, DeFrancesco TC,

537 Lehmkuhl LB, Kellihan HB, Bulmer B, Gordon SG, Cunningham SM, MacGregor J, Stepien

- 538 RL, Lefbom B, Adin D, Lamb K. Relationship of plasma N-terminal pro-brain natriuretic
- 539 peptide concentrations to heart failure classification and cause of respiratory distress in dogs
- using a 2nd generation ELISA assay. J Vet Intern Med. 2015;29:171-9.
- 541 19. Hansson K, Haggstrom J, Kvart C, Lord P. Left atrial to aortic root indices using two-
- 542 dimensional and M-mode echocardiography in cavalier King Charles spaniels with and
- 543 without left atrial enlargement. Vet Radiol Ultrasound. 2002;43:568-75.

544 20. Cornell CC, Kittleson MD, Della Torre P, Haggstrom J, Lombard CW, Pedersen HD,
545 Vollmar A, Wey A. Allometric scaling of M-mode cardiac measurements in normal adult
546 dogs. J Vet Intern Med. 2004;18:311-21.

54721.Schulz KF, Altman DG, Moher D, Group C. CONSORT 2010 statement: updated

548 guidelines for reporting parallel group randomized trials. Obstet Gynecol. 2010;115:1063-70.

549 22. Hezzell MJ, Boswood A, Moonarmart W, Elliott J. Selected echocardiographic
550 variables change more rapidly in dogs that die from myxomatous mitral valve disease. J Vet
551 Cardiol. 2012;14:269-79.

Lord PF, Hansson K, Carnabuci C, Kvart C, Haggstrom J. Radiographic heart size
and its rate of increase as tests for onset of congestive heart failure in Cavalier King Charles
Spaniels with mitral valve regurgitation. J Vet Intern Med. 2011;25:1312-9.

555 24. Stowasser M, Ahmed AH, Pimenta E, Taylor PJ, Gordon RD. Factors affecting the
aldosterone/renin ratio. Horm Metab Res. 2012;44:170-6.

557 25. Lefebvre HP, Ollivier E, Atkins CE, Combes B, Concordet D, Kaltsatos V, Baduel L.

Safety of spironolactone in dogs with chronic heart failure because of degenerative valvular
disease: a population-based, longitudinal study. J Vet Intern Med. 2013;27:1083-91.

560 26. Bonagura JD, Schober KE. Can ventricular function be assessed by echocardiography
561 in chronic canine mitral valve disease? J Small Anim Pract. 2009;50 Suppl 1:12-24.

562 27. Schober KE, Cornand C, Kirbach B, Aupperle H, Oechtering G. Serum cardiac

troponin I and cardiac troponin T concentrations in dogs with gastric dilatation-volvulus. J

564 Am Vet Med Assoc. 2002;221:381-8.

28. Langhorn R, Willesen JL. Cardiac Troponins in Dogs and Cats. J Vet Intern Med.
2016;30:36-50.

- 567 29. Guglin M, Kristof-Kuteyeva O, Novotorova I, Pratap P. Aldosterone antagonists in
- heart failure. J Cardiovasc Pharmacol Ther. 2011;16:150-9.

- 570 Figure 1: Summary flow diagram detailing the tests performed at each visit during the study
- 571 period.



- 586 ECG, electrocardiogram; PCV, packed cell volume; NT-proBNP, N-terminal B-type
- 587 natriuretic peptide; cTnI, cardiac troponin I.

588

Figure 2: Flow diagram of the progress through the phases of this parallel, randomized trial oftwo groups. Analysis was on the basis of intention to treat.



Figure 3: LA/Ao in dogs receiving placebo and those receiving spironolactone at baseline (n = 12 and 13, respectively) and at the 6 month time point (n = 12 and 12, respectively). The central tendency represents the mean and the error bars represent the standard deviation.



594

595 Figure 4: LVIDDN (cm/ [kg^{0.294}]) in dogs receiving placebo and those receiving

spironolactone at baseline (n = 12 and 13, respectively) and at the 6 month time point (n = 12
and 12, respectively). The central tendency represents the mean and the error bars represent
the standard deviation.



Figure 5: Serum NT-proBNP concentrations (pmol/L) in dogs receiving placebo and those receiving spironolactone at baseline (n = 12 and 13, respectively) and at the 6 month time point (n = 12 and 11, respectively). The central tendency represents the mean and the error bars represent the standard deviation.



605

606

608 receiving placebo and those receiving spironolactone at baseline.

Variable	Group receiving	Group receiving	P value
	placebo	spironolactone	
	(n = 12)	(n = 13)	
Age (years)	9.4 ±2.0	9.7 ±2.0	0.707
CKCS (yes/ total)	7/ 12	9/ 13	0.688
Sex (male/ total)	7/ 12	11/13	0.202
Body weight (kg)	11.6 ±4.1	11.8 ±6.3	0.940
Systolic blood pressure	156.5 ±29.8	159.4 ±32.0	0.816
(mmHg)			
Heart rate (ECG) (bpm)	117.3 ±25.0	117.2 ±25.7	0.992
LA:Ao ratio	1.27 ±0.17	1.40 ±0.21	0.105
LVIDdN (cm/[kg ^{0.294}])	1.79 ±0.20	1.95 ±0.35	0.180
LVIDsN (cm/[kg ^{0.315}])	1.05 ±0.13	1.10 ±0.28	0.548
LVIDd/ LVFWd ratio	4.06 ±0.63	4.46 ±0.96	0.229
E wave velocity (m/s)	1.01 ±0.22	0.99 ±0.35	0.883
E/ A wave ratio	1.37 ±0.22	1.40 ±0.50	0.857
Serum NT-proBNP (pmol/L)	1006.0	2434.0	0.005
	[541.0, 2108.0]	[740.0, 3955.0]	
Serum cTnI (ng/mL)	0.03	0.03	0.437
	[0.01, 0.16]	[0.01, 0.19]	
UAC (g/mol)	0.079	0.088	0.744
	[0.044, 0.119]	[0.042, 0.165]	
PCV (%)	42.7 ±6.4	40.9 ±4.6	0.439
Serum urea (mmol/L)	5.3 ±2.2	6.0 ±1.9	0.426

Serum creatinine (µmol/L)	68.9 [59.7, 146.3]	70.5 [52.5, 130.6]	0.810
Serum Na ⁺ (mmol/L)	147.7 ±2.5	147.8 ±2.0	0.893
Serum K ⁺ (mmol/L)	4.2 ±0.4	4.5 ±0.5	0.078
Serum Cl ⁻ (mmol/L)	111.5 ± 3.6	113.0 ±2.5	0.237

All variables in the group receiving placebo included 12 dogs, except UAC (n = 10), E wave 610 velocity (n = 11) and E/A ratio (n = 11). All variables in the group receiving spironolactone 611 included 13 dogs, except UAC (n = 9). The P values for variables for which significant 612 between-group differences were detected (P < 0.05) are highlighted in bold text. 613 CKCS, cavalier King Charles spaniel; ECG, electrocardiogram; bpm, beats per minute; 614 615 LA:Ao ratio, left atrial to aortic root ratio; LVIDdN, left ventricular end-diastolic diameter normalized for body weight; LVIDsN, left ventricular end-systolic diameter normalized for 616 617 body weight; LVIDd/ LVFWd ratio, left ventricular end-diastolic diameter to left ventricular free wall thickness in diastole ratio; NT-proBNP, N-terminal B-type pro-natriuretic peptide; 618 cTnI, cardiac troponin I; UAC, urinary aldosterone to creatinine ratio; PCV, packed cell 619

620 volume; Na^+ , sodium ions; K^+ , potassium ions; Cl^- , chloride ions.

- 621 Table 2: Repeated measures linear mixed model analysis of change over 6 months in
- 622 variables of primary interest.

Variable	P (between	Unit change in variable per	SE	P (within
	groups)	month (B)		group)
		[95% confidence interval]		
Serum NT-proBNP (pmol/L)	0.087			
Placebo group ($n = 12$)		54.93 [-5.58 to 115.45]	29.22	0.073
Spironolactone group $(n = 12)$		-18.50 [-78.23 to 41.24]	29.00	0.529
Log (Serum cTnI [ng/mL])	0.708			
Placebo group (n = 12)		-0.010 [-0.035 to 0.015]	0.012	0.420
Spironolactone group ($n = 12$)		-0.004 [-0.028 to 0.021]	0.012	0.772
LA:Ao ratio	0.110			-
Placebo group ($n = 12$)		0.020 [0.008 to 0.032]	0.006	0.002
Spironolactone group (n = 12)		0.007 [-0.005 to 0.018]	0.006	0.231
0.294 LVIDdN (cm/[kg])	0.223			
Placebo group ($n = 12$)		0.017 [0.006 to 0.029]	0.006	0.005
Spironolactone group ($n = 12$)		0.007 [-0.004 to 0.019]	0.006	0.194
	1		1	1

- 624 Repeated measures linear mixed model analysis of change over 6 months in variables of
- primary interest (LA:Ao ratio, LVIDdN and serum NT-proBNP and cTnI) for the verum andplacebo groups.
- 627 P (between groups): probability that the rate of change of the variable is different between the
- 628 group receiving placebo and the group receiving spironolactone.

Table 3: Repeated measures linear mixed model analysis of change over 6 months in clinical,

630 echocardiographic and biomarker parameters of secondary interest.

Variable	P (between	Unit change in variable per	SE	P (within
	groups)	month (B)		group)
		[95% confidence interval]		
PCV (%)	0.792			
Placebo group $(n = 9)$		-0.06 [-0.41 to 0.29]	0.17	0.729
Spironolactone group (n = 10)		0.01 [-0.35 to 0.36]	0.17	0.977
Serum urea (mmol/L)	0.406			
Placebo group ($n = 12$)		0.01 [-0.15 to 0.17]	0.08	0.903
Spironolactone group (n = 12)		0.11 [-0.06 to 0.27]	0.08	0.195
Log (Serum creatinine [µmol/L])	0.223			-
Placebo group $(n = 12)$				
Spinon alextens group $(n - 12)$		-0.001 [-0.006 to 0.004]	0.002	0.706
Spironolactone group ($n = 12$)		0.003 [-0.002 to 0.008]	0.002	0.177
Serum Na ⁺ (mmol/L)	0.407			
Placebo group ($n = 12$)		-0.071 [-0.254 to 0.113]	0.091	0.441
Spironolactone group (n = 12)		0.036 [-0.144 to 0.216]	0.089	0.690
Serum K ⁺ (mmol/L)	0.328			
Placebo group ($n = 12$)		0.028 [-0.011 to 0.067]	0.019	0.157
Spironolactone group (n = 12)		0.001 [-0.037 to 0.039]	0.019	0.957
Serum Cl ⁻ (mmol/L)	0.713			
Placebo group $(n = 12)$		0.023 [-0.258 to 0.306]	0.139	0.871

Spironolactone group $(n = 12)$		-0.050 [-0.327 to 0.228]	0.138	0.719
Log (UAC[g/mol])	0.223			
Placebo group $(n = 0)$		0 017 [0 006 to 0 029]	0.006	0.005
Tracebo group (II – 9)		0.017 [0.000 to 0.029]	0.000	0.005
Spironolactone group $(n = 8)$		0.007 [-0.004 to 0.019]	0.006	0.194
	0.456			
Body weight (kg)	0.456			
Placebo group $(n = 12)$		-0.016 [-0.055 to 0.024]	0.019	0.428
			0.010	0.044
Spironolactone group $(n = 12)$		-0.036 [-0.074 to 0.002]	0.019	0.066
LVIDsN (cm/[kg ^{0.315}])	0.839			
Placebo group $(n = 12)$		0.005 [-0.006 to 0.015]	0.005	0.367
Spironolactone group $(n = 12)$		0.003 [-0.007 to 0.013]	0.005	0.527
LVIDd/ LVFWd ratio	0.531			
Placebo group $(n = 12)$		0 053 [-0 005 to 0 029]	0.028	0.070
		0.000 [0.000 to 0.027]	0.020	0.070
Spironolactone group $(n = 12)$		0.028 [-0.029 to 0.086]	0.028	0.315
	0.724			
Log (E wave velocity [m/s])	0./34			
Placebo group $(n = 11)$		0.001 [-0.006 to 0.009]	0.004	0.740
		0.001 5.0.000 / 0.0071	0.002	0.007
Spironolactone group ($n=12$)		-0.001 [-0.008 to 0.007]	0.003	0.887
E/A ratio	0.876			
			0.01.5	
Placebo group (n =11)		-0.004 [-0.034 to 0.026]	0.015	0.804
Spironolactone group $(n = 12)$		-0.007 [-0.035 to 0.022]	0.014	0.625
Heart rate (ECG) (bpm)	0.477			
Placebo group $(n = 12)$		-0.066 [-1.684 to 1.553]	0.781	0.934
		- *		
Spironolactone group $(n = 12)$		-0.858 [-2.449 to 0.733]	0.768	0.276

632 Repeated measures linear mixed model analysis of change over 6 months in clinical,

633 echocardiographic and biomarker parameters of secondary interest for the verum and placebo

634 groups.

- 635 P (between groups): probability that the rate of change of the variable is different between the
- 636 group receiving placebo and the group receiving spironolactone.
- 637 P (within group): probability that the rate of change over time within the group = 0.
- 638 SE, standard error; for definitions of other abbreviations see legend to Table 1.

- Table 4: Comparisons of selected clinical, echocardiographic and biomarker data between
- 640 dogs receiving placebo and those receiving spironolactone at the 2 week time point.
- 641

Variable	Group receiving placebo	Group receiving	P value
	(n = 10)	spironolactone	
		(n = 10)	
UAC (g/mol)	0.063 [0.032, 0.077]	0.236 [0.125, 0.339]	0.006
PCV (%)	46.0 ±7.8	42.9 ±7.4	0.428
Serum urea (mmol/L)	6.3 ±2.2	6.4 ±2.0	0.925
Serum creatinine (µmol/L)	74.7 [63.4, 93.2]	76.4 [67.7, 83.7]	0.940
Serum Na ⁺ (mmol/L)	147.1 ±1.7	147.3 ±1.1	0.780
Serum K ⁺ (mmol/L)	4.5 ±0.2	4.7 ±0.2	0.098
Serum Cl ⁻ (mmol/L)	109.3 ±2.8	111.4 ±3.2	0.144

642 Comparisons of selected clinical, echocardiographic and biomarker data between dogs

643 receiving placebo and those receiving spironolactone at the 2 week time point. All variables

644 in the group receiving placebo included 10 dogs, except PCV and UAC, which each included

645 7 dogs. All variables in the group receiving spironolactone included 10 dogs, except PCV,

646 which included 9 dogs and UAC, which included 7 dogs. The P values for variables for

647 which significant between-group differences were detected (P < 0.05) are highlighted in bold

648 text.

649 For definitions of abbreviations see legend to Table 1.

651 receiving placebo and those receiving spironolactone at the 6 month time point.

Variable	Group receiving placebo (n	Group receiving	P value
	= 12)	spironolactone ($n = 12$)	
Body weight (kg)	11.5 ±4.2	11.5 ±6.9	0.986
Heart rate (ECG) (bpm)	116.8 ±25.8	111.7 ±20.4	0.592
LA:Ao ratio	1.39 ±0.25	1.46 ±0.18	0.427
LVIDdN (cm/[kg ^{0.294}])	1.89 ±0.24	2.01 ±0.34	0.367
LVIDsN (cm/[kg ^{0.315}])	1.08 ±0.18	1.11 ±0.28	0.747
LVIDd/ LVFWd ratio	4.37 ±0.82	4.59 ±1.12	0.593
E wave velocity (m/s)	1.02 ±0.19	0.96 ±0.28	0.539
E/ A wave ratio	1.33 ±0.13	1.37 ±0.39	0.722
NT-proBNP (pmol/L)	.1188.5 [359.0, 3018.0]	1852.0 [765.0, 4128.0]	0.051
cTnI (ng/mL)	0.025 [0.010, 0.150]	0.030 [0.010, 0.170]	0.378
UAC (g/mol)	0.063 [0.010, 0.410]	0.125 [0.060, 0.370]	0.113
PCV (%)	42.8±6.3	42.1 ±5.5	0.792
Serum urea (mmol/ L)	5.7 ±1.7	6.7 ±3.5	0.364
Serum creatinine (µmol/L)	76.1 [51.9, 146.3]	82.4 [54.1, 145.5]	0.514
Serum Na ⁺ (mmol/L)	147.1 ±1.0	147.9 ±1.4	0.127
Serum K ⁺ (mmol/L)	4.5 ±0.3	4.6 ±0.3	0.448

	Serum Cl ⁻ (mmol/L)	111.0 ±2.6	112.5 ±3.6	0.258	
652	Comparisons of clinical, echocar	rdiographic and biomarker	data between dogs receiving	ing	
653	placebo and those receiving spir	onolactone at the 6 month	time point. All variables in	n the	
654	group receiving placebo included 12 dogs, except UAC ($n = 9$). All variables in the group				
655	receiving spironolactone include	ed 12 dogs, except PCV (n	= 11), NT-proBNP (n = 1	1) and	
656	UAC (n =6).				

657 For definitions of abbreviations see legend to Table 1.