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1	Extracorporeal Liver Support Devices for Listed Patients
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12	Keywords (5 not in title): albumin dialysis, bioartificial liver, acute liver failure, acute-on-

13 chronic liver failure, plasma exchange

1 Abbreviations: ACLF, acute-on-chronic liver failure; ALF, acute liver failure; AMC-BAL, 2 Academic Medical Centre Bioartificial Liver; ELAD™, Extracorporeal Liver Assist Device™; 3 ELSD, extracorporeal liver support device; HE, hepatic encephalopathy; HSA, human serum albumin; HVP, high-volume therapeutic plasma exchange; ITT, intention to treat; Li-ALS, Li-4 Artificial Liver Support; MARS<sup>®</sup>, molecular adsorbents recirculating system; n.s., no 5 6 significant difference between groups; PP, per protocol; SMT, standard medical therapy; 7 SPAD<sup>®</sup>, single pass albumin dialysis; SRBAL, Spheroid Reservoir Bioartificial Liver; TPE, therapeutic plasma exchange; UCL-LDD, University College London-Liver Dialysis Device. 8 9 **Conflicts of Interest:** The authors have nothing to declare with the following exceptions. 10 Rajiv Jalan has research collaborations with Ocera, Grifols, Norgine and Gambro, consults 11 12 for Ocera and Conatus and has received speaking fees from Norgine and Grifols. Rajiv 13 Jalan is the inventor of University College London-Liver Dialysis Device, which has been 14 patented by UCL and licensed to Yagrit Limited. 15 \*Corresponding author: Professor Rajiv Jalan, MD PhD. Liver Failure Group, Institute of 16 17 Liver and Digestive Health, University College London Medical School Royal Free Campus, Roland Hill Street, London, NW3 2PF, UK. Tel: +442074332795. Fax: +442074332775. 18

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#### 1 ABSTRACT

2 An alternative to liver transplantation for patients with liver failure remains an unmet need. In 3 acute liver failure, the ideal extracorporeal liver support device would replace the functions of 4 the failing liver in order to permit spontaneous recovery, given the incredible regenerative 5 potential of the liver, negating the need for transplantation. In acute-on-chronic liver failure, 6 an extracorporeal liver support device would ideally support hepatic function until recovery to 7 liver function prior to acute decompensation or until liver transplantation. In decompensated 8 cirrhosis, an extracorporeal liver support device could again be used to support hepatic 9 function until transplant. In addition, extracorporeal liver support devices may have potential 10 to treat the multi-organ failure that accompanies liver failure including hepatic 11 encephalopathy, renal failure and immune dysfunction or indeed potential to promote liver 12 regeneration. Creation of an extracorporeal bioartificial liver able to completely replace liver 13 function remains an unmet need. This review will describe a number of technologies suitable for clinical trials in man, which have resulted from decades of engineering and biological 14 15 research to develop a bioreactor able to adequately sustain functional hepatocytes. In addition, this review will describe artificial liver support devices, primarily designed to replace 16 17 the detoxifying functions of the liver and consider the current data available or studies required to support their use in liver failure patients on the transplant waiting list. 18

19

#### 20 INTRODUCTION

Mortality in patients with liver failure who cannot be rescued with liver transplantation 21 remains high despite improvements in supportive care (1). The fundamental thinking behind 22 the use of extracorporeal liver support devices (ELSD) is the idea that if the patient's liver 23 and extrahepatic organs can be supported long enough, recovery should be possible, 24 because of the regeneration potential of the liver (2). Alternative aims of ELSD may be to 25 'bridge' liver failure patients to liver transplantation or to support patients with end-stage liver 26 disease while on the waiting list for transplantation. Additional therapeutic goals may be to 27 28 treat end organ dysfunction such as hepatic encephalopathy (HE), renal failure or immune

dysfunction (3). Finally, as one understands the pathophysiological basis of regeneration or
its inhibition better, ELSD may be used to target particular molecules to enhance this
regenerative process. Depending upon whether the liver failure occurs on the background of
a previously healthy liver or in patients with underlying chronic liver disease, the conditions
are referred to as acute liver failure (ALF) or acute-on-chronic liver failure (ACLF)
respectively (Table 1).

7

ALF is a rare disease and is defined as the occurrence of HE in patients with severe acute liver injury within 6-months of the onset of symptoms (4). From a pathophysiological perspective, patients with ALF are the perfect group of patients likely to benefit from ELSD because recovery is likely to return the patient to their pre-liver failure state, in which there is no pre-existing liver pathology. It is becoming clear that in addition to providing support for hepatic function, modulation of hepatic and systemic inflammation will be important to prevent deaths either from an exaggerated inflammatory response or infection (3).

15

16 ACLF is much more common than ALF and typically occurs in patients with cirrhosis (5). The 17 condition is characterized by acute deterioration of a cirrhotic patient with or without a recognized precipitating event, associated with organ failures and high mortality rates (6). 18 19 Data from prospective studies are now available that allow accurate, sequential assessments of patients, which provide prognostic information. The CLIF Consortium organ 20 failure score is used for diagnosis of the syndrome (Table 2) and the CLIF Consortium ACLF 21 score for defining the prognosis (7). A pre-ACLF group has now been identified, which will 22 allow studies of ELSD to prevent the occurrence of ACLF in susceptible patients (8). 23 Systemic inflammation is the key pathophysiological factor that drives the syndrome making 24 this a particular target of ELSDs (9). The aim of ELSDs in patients with ACLF is to support 25 hepatic function during acute decompensation until recovery to baseline liver function and/or 26 27 liver transplantation.

28

Decompensated cirrhosis is pathophysiologically different and typically represents patients
 that have end-stage cirrhosis with varying degrees of end-organ dysfunction. In this group of
 patients, ELSD is aimed at supporting them until liver transplantation.

4

5 This review describes the state of the art about the types of ELSDs that are available, the 6 results of the large and important clinical trials and the new ELSDs that are in or about to 7 enter clinical trials. The reported human, randomised, controlled, clinical trials of ELSDs, for 8 which survival was the primary outcome, are given in Table 3 with selected survival data 9 shown in Figures 1 to 3.

10

## 11 CURRENTLY AVAILABLE EXTRACORPOREAL LIVER SUPPORT DEVICES

The currently available artificial ELSDs are based on the principal of removal of protein bound and water soluble substances (blood purification) by albumin dialysis, by plasma separation and filtration or by therapeutic plasma exchange. Devices based solely on the removal of water soluble substances (blood detoxification) have not shown any benefit in survival, possibly because of the limited, non-specific absorptive capacity of chemical adsorbents (10).

18

#### 19 The following artificial ELSDs are currently available:

20	(i)	The Molecular Adsorbents Recirculating System (MARS®, Gambro, Sweden) was
21		first described in 1993 (Supplementary Material Figure S1) (11). In $MARS^{\mathbb{8}}$ , blood
22		is dialyzed across an albumin-impermeable, approximately 50-60 kDa cut-off,
23		membrane against 20% human serum albumin (HSA). HSA solution is
24		continuously stripped of protein bound and water soluble toxins by passage
25		through a secondary circuit containing a charcoal column, an anion exchange
26		resin column and a low-flux dialyzer (12-14).
27	(ii)	The Fractionated Plasma Separation, Adsorption and Dialysis device

28 (Prometheus<sup>®</sup>, Fresenius Medical Care, Germany) separates the patient's

1		albumin/plasma from blood by passage across an approximately 300 kDa cut-off
2		membrane (Supplementary Material Figure S2). Patient albumin/plasma is then
3		passed directly over two columns containing different adsorbents. A high-flux
4		dialyzer inserted into the blood circuit clears water-soluble substances (15, 16).
5	(iii)	Single pass albumin dialysis® (SPAD®) can be carried out with a standard dialysis
6		setup, by use of hollow fibres made of a high-flux albumin-impermeable
7		membrane and the addition of HSA to the dialysis solution to enable solute
8		transfer from the patient's blood to the dialysis solution (Supplementary Material
9		Figure S3) (17, 18).
10	(iv)	Therapeutic plasma exchange (TPE) involves extracorporeal separation and
11		removal of patient plasma from blood and return of blood cells with a replacement
12		fluid to the patient. Fresh frozen plasma is the typical replacement fluid, but HSA
13		has also been reported (19).
14		
15	MARS®, F	Prometheus <sup>®</sup> and SPAD <sup>®</sup> are all able to reduce serum bilirubin and bile acids.
16	Studies co	omparing MARS <sup>®</sup> and Prometheus <sup>®</sup> in ACLF show higher efficiency of
17	Promethe	$us^{\circ}$ for removal of bilirubin and urea and equal efficiency for removal of bile acids
18	(20, 21). H	However, an actual improvement of synthetic liver function has neither been
19	expected	nor observed. For patients awaiting liver transplantation improvement of systemic
20	haemody	namics, renal function or HE might be able to "buy" valuable time until an organ
21	becomes	available, serve as a bridge to recovery and it can be hypothesized that this would
22	also impa	ct on prognosis after transplantation.
23		
24	Molecula	r Adsorbents Recirculating System <sup>®</sup>
25	A meta-ar	nalysis (22) of 4 randomized (14, 23-25) and 2 selected non-randomized trials (26,
26	27) did no	ot show any overall effect of MARS <sup>®</sup> on mortality. However, explorative analysis of

as compared to the standard medical treatment (SMT) group (22). Another randomized

controlled trial showed that MARS® therapy in patients with ACLF has a beneficial effect on 1 circulating neurohormones, nitric oxide and free radical production, and reduces markers of 2 3 oxidative stress (28). The clinical effects of these changes are reflected in individual organ function with temporal improvement in cholestasis, liver function, renal function, 4 5 encephalopathy, and in some patients, mean arterial pressure (28). Indeed, one of the most consistent findings in studies of MARS<sup>®</sup> in ACLF is an improvement in portal and systemic 6 7 haemodynamics (29-31). Furthermore a large randomized controlled trial revealed a significant effect of MARS<sup>®</sup> on the severity of HE (32). The largest study so far – the RELIEF 8 trial – however could not show a benefit of MARS® on mortality in ACLF, but demonstrated 9 10 safety, a dialysis effect and a modest effect on HE (33) (Figure 1, Table 3). Failure to show a 11 survival benefit may have been due to the heterogeneous patient population. However, 12 another large, randomized study in ALF – the FULMAR trial – also failed to show a survival 13 benefit of MARS<sup>®</sup> (Table 3). In this study most patients were transplanted within a median of 16.2 hours, leaving little time for a liver support system to demonstrate its effect (34). A 14 retrospective study of continuous MARS® treatment in critically ill patients listed for liver 15 transplantation with ALF, ACLF or graft dysfunction, showed that MARS<sup>®</sup> may be of value as 16 17 a bridge to transplant but also revealed severe side effects with respect to coagulation and electrolytes (35). Therefore, the use of MARS<sup>®</sup> in patients with liver failure waiting for an 18 19 organ should be performed under close observation with treatment of coagulopathy and electrolyte disturbances (35). In another single centre observation from the Netherlands that 20 included 20 children with ALF or graft dysfunction, MARS® could be successfully applied, but 21 with similar coagulation side effects and the need for liver transplantation was not reduced 22 (36). Another single centre experience from Mexico suggested that MARS<sup>®</sup> reduced the 23 need for liver transplantation by contributing to native liver recovery (37). However, a 24 retrospective cohort study is not the optimal study design to answer this question. 25

26

From the available data, it is not possible to conclude whether or not MARS<sup>®</sup> is beneficial for
patients on the transplant waiting list. It is possible that efficiency of the device is not optimal.

- Therefore, the development of a device with higher efficiency might be of value. Recently,
   the use of a double absorption unit in parallel has been tested (38).
- 3

#### 4 **Prometheus**<sup>®</sup>

5 Initial and subsequent uncontrolled data for Prometheus<sup>®</sup> show high elimination of albumin bound toxins and good safety data (16, 39). Comparable to MARS<sup>®</sup>, Prometheus<sup>®</sup> can be 6 7 used safely in patients awaiting an urgent liver transplantation (40), but severe coagulation disturbances have been reported (41). The largest cohort study of Prometheus® in ALF 8 9 patients was performed in Turkey and demonstrated safety and efficacy: one third of patients survived without transplantation, leading the authors to suggest that Prometheus<sup>®</sup> may be 10 effective as a bridge to recovery (42). However, as for MARS<sup>®</sup>, Prometheus<sup>®</sup> failed to 11 12 improve survival of patients with ACLF in a large prospective randomized study (Figure 2, Table 3) (43). Therefore the current data does not allow us to conclude whether or not 13 Prometheus<sup>®</sup> is of benefit to patients on the waiting list. 14

15

## 16 Single pass albumin dialysis<sup>®</sup>

SPAD<sup>®</sup> can be used with any dialysis setup, therefore there is no need to invest in an extra 17 machine. However, the amount of HSA required is high. SPAD® has been mainly tested in 18 19 vitro and reported in case reports. In vitro there is evidence that the detoxification capacity of 20 SPAD<sup>®</sup> is greater than MARS<sup>®</sup>(18). In a retrospective study, MARS<sup>®</sup> and SPAD<sup>®</sup> showed equal efficacy (44). In ALF, SPAD® was well tolerated but failed to improve survival and did 21 not change referral to liver transplantation (45). In a single-center experience from Germany, 22 SPAD® did not have any impact on survival or transplantation rate in patients with ACLF 23 listed for transplantation (46). However, patient numbers were small. Again, it is not possible 24 to conclude whether or not SPAD® is beneficial for patients on the waiting list. 25

26

## 27 Therapeutic plasma exchange

1 Therapeutic plasma exchange has been reported in isolated ALF and ACLF patients since the 1960s. Rationale has been removal of all toxins, as well as harmful inflammatory 2 3 mediators and replacement of beneficial plasma proteins normally synthesised by the liver. 4 In liver failure, TPE has been shown to reduce serum bilirubin and ammonia and to increase 5 coagulation factors improving coagulopathy. Hypocalcaemia and alkalosis occur due to 6 anticoagulant use, but are easily corrected (19). A recent multi-centre open randomised controlled trial of high-volume TPE (HVP, exchange of approximately 8-12 litres or 15% body 7 8 weight of plasma), on three consecutive days, in 182 ALF patients demonstrated increased 9 survival to hospital discharge (Figure 3, Table 3) (47). In patients who fulfilled poor 10 prognostic criteria, but were not listed for transplant, HVP (n=28) increased survival 11 compared to SMT (n=36). This survival advantage was associated with immune modulation and improvement in renal function, cardiovascular status, SOFA score and CLIF-SOFA 12 13 score (47).

14

## 15 EMERGING TECHNOLOGIES IN EXTRACORPOREAL LIVER SUPPORT DEVICES

Bioartificial ELSDs include a bioreactor that contains hepatocytes, which in the most ideal 16 17 scenario, would replace the functions of the failing liver including: ammonia detoxification via the urea cycle; drug metabolism; protein synthesis; and carbohydrate and lipid metabolism 18 (48). Bioartificial ELSD development has been limited by their requirement for primary 19 hepatocytes, which demonstrate better hepatocyte functionality compared to immortalised 20 cell lines, but with the accompanying disadvantage of reduced cell viability and limited 21 availability (48). Moreover, bioreactor design has been challenged with maintaining large 22 hepatocyte cultures for effective patient treatment, whilst simultaneously acting as an 23 24 effective interface between bioreactor hepatocyte function and patient plasma (48). 25 Nevertheless progressive evolution of bioreactor design and hepatocyte biology has resulted in bioreactors with considerable hope for ALF and ACLF treatment. These include: 26 Extracorporeal Liver Assist Device<sup>™</sup> (ELAD<sup>™</sup>); Academic Medical Centre Bioartificial Liver 27 (AMC-BAL); and Spheroid Reservoir Bioartificial Liver (SRBAL). ELAD has entered human 28

- clinical trials. SRBAL and the latest version of AMC-BAL have shown efficacy in animal
   experiments, but data from human clinical trials are currently unavailable.
- 3

A number of artificial ELSDs are in development that may either improve detoxification
compared to current ELSDs or combine detoxification with techniques to attenuate liver
injury. These include: Hepa Wash®; Li-Artificial Liver Support (49) and University College
London-Liver Dialysis Device (UCL-LDD). All three of these devices have shown efficacy in
animal experiments, but data from human clinical trials are currently unavailable.

9

## 10 Extracorporeal Liver Assist Device

11 ELAD<sup>™</sup> has been trialled in animal models of ALF and human liver failure patients since the 12 1990s (50-52). Its key component is a guartet of hollow fibre dialysis cartridges containing 13 HepG2/C3A cells, a human hepatoblastoma cell line, within the extra-fibre spaces (Supplementary Material Figure S4). HepG2/C3A cells remain viable throughout the 14 15 recommended 3-10 day treatment (53). HepG2/C3A cells demonstrate albumin synthesis 16 and cytochrome P450 activity, but functionality is significantly less than primary hepatocytes 17 with failure to detoxify ammonia via the urea cycle (48, 54). Early phase I pilot studies in limited numbers of human ALF patients have demonstrated safety, but no improvement in 18 survival and biochemical and clinical parameters (51, 52). Preliminary results of a trial in 19 patients with acute decompensation of chronic hepatitis B or C reported significant extension 20 of 30 day transplant free survival and biochemical improvement (Table 3) (55). Clinical trials 21 of ELAD<sup>™</sup> in ACLF, ALF, severe acute alcoholic hepatitis and alcoholic-induced liver failure 22 are currently ongoing (56). In a recent press release, the results of the large randomised trial 23 of ELAD<sup>™</sup> in alcohol-related ACLF patients were reported to be negative (57). The full report 24 is awaited. 25

26

## 27 Academic Medical Centre Bioartificial Liver

1 The AMC-BAL has been in development since the 1990s (58). Key bioreactor features are: a non-woven matrix for 3-D hepatocyte cultures; spiralling of this 3-D matrix around oxygen 2 3 carrying capillaries; and direct exposure of hepatocytes to patient plasma (Supplementary 4 Material Figure S5). Primary hepatocyte viability has been reported to be 90% on day three. 5 The first phase I clinical trial of AMC-BAL in man used a device containing primary porcine 6 hepatocytes. In this trial 12 ALF patients were treated for 4 to 35h: eleven were successfully 7 bridged to liver transplantation and one recovered spontaneously. AMC-BAL treatment was 8 associated with improvement in neurological and haemodynamic status in all patients; 9 improvement in renal function in those with renal insufficiency and reduction in 10 hyperbilirubinaemia and lactic acidosis (58). Porcine endogenous retrovirus DNA was found 11 in patient plasma directly after treatment, but was undetectable thereafter. Nevertheless 12 clinical use of this device was restricted due to ethical, immunological and zoonotic 13 concerns.

14

Recently the HepaRG human hepatoma cell line has been cultured in the AMC-BAL instead 15 16 of primary porcine hepatocytes. HepaRG cells approximate primary hepatocyte cultures 17 more than any other human hepatocyte cell line (48). Culture within the AMC-BAL: 1) increased hepatic functionality with respect to ammonia elimination, the urea cycle and 18 19 cytochrome P450 activity and 2) revealed lactate consumption, amino acid metabolism, drug metabolism and bile acid production similar to that of primary hepatocytes (59). In a rat ALF 20 model, the HepaRG-AMC-BAL resulted in a 50% increase in survival and delay in 21 progression of HE, kidney failure and hyperammonaemia (60). 22 23

## 24 Spheroid Reservoir Bioartificial Liver (SRBAL)

SRBAL has been in development since the early 2000s. Its key component is a bioreactor
containing primary porcine hepatocytes in suspension, which when exposed to an oscillation
frequency of 0.25Hz cluster into spheroids with stable cell viability (Supplementary Material
Figure S6) (61, 62). Hepatocyte spheroids demonstrate good hepatocyte function in terms

1 of: phase I and phase II drug metabolism; ammonia conversion to urea via the urea cycle;

2 and albumin synthesis (61). A trial using a pig ALF model has been reported (63). Pigs were

3 treated either with two 6-hour treatments (intermittent) or one 24 hour treatment

4 (continuous). Both SRBAL treatments improved survival and reduced hyperammonaemia

5 and continuous SRBAL reduced intracranial hypertension and brain water.

6

## 7 Hepa Wash<sup>®</sup>

Hepa Wash<sup>®</sup> is an artificial ELSD that detoxifies blood by albumin dialysis against a 2%
albumin dialysate (64). The albumin dialysate is recirculated via a 'Hepa Wash' circuit, which
contains two parallel conventional haemofilters, in which albumin bound toxins are released
through exposure to an alkaline or acid environment and subsequently removed by filtration.
This design aims to maintain clearances of protein bound toxins through the treatment
period (64). This is contrary to MARS<sup>®</sup>, where a decline in clearance of protein bound toxins

15

16 In a pig liver ischaemia ALF model, Hepa Wash<sup>®</sup> resulted in improvement in survival,

17 cerebral perfusion pressure, haemodynamic status and kidney function. Moreover, Hepa

18 Wash® resulted in reduction in azotaemia, hyperammonaemia, and blood nitrate/nitrite

19 levels (64). Clinical trials in humans with ALF and ACLF were initiated in 2010, but have

since been terminated for unknown reasons (NCT01079104, NCT01079091).

21

## 22 Li-Artificial Liver Support (Li-ALS)

Li-ALS is an artificial ELSD that combines a low-volume TPE (exchange of approximately
2.5% body weight of plasma) circuit with a modified MARS secondary circuit, in which highflux hemofiltration replaces low-flux haemodialysis (49). This approach seeks to benefit from
the more comprehensive detoxification achieved by TPE compared to MARS, without need
for a supply of exogenous fresh frozen plasma, as patient plasma is returned postdetoxification to the patient. In a D-galactosamine pig model of ALF, Li-ALS resulted in an

improvement in survival compared to treatment with low-volume TPE alone and to treatment
 with the modified MARS circuit alone (49).

3

#### 4 University College London-Liver Dialysis Device (UCL-LDD)

5 UCL-LDD is an artificial ELSD, in which blood is filtered across a high-cut off membrane 6 (nominal cut-off of 60kDa) and then passed over a selective endotoxin adsorption 7 membrane. Filtration across a high-cut off membrane results in albumin loss, which is 8 replaced by HSA infusion (66). The resultant albumin exchange is proposed to correct 9 irreversible loss of detoxifying function of albumin reported in liver failure. Reduction in 10 endotoxaemia aims to reduce innate immune response, which worsens liver injury. Moreover 11 high-cut off filters reduce circulating pro- and anti-inflammatory cytokines and correct 12 immune dysfunction in septic patients with acute renal failure (67), so the same may apply to 13 ALF. In a pig model of paracetamol-induced ALF, UCL-LDD improved survival and cardiovascular and respiratory function and reduced circulating dysfunctional albumin, 14 endotoxaemia and immune system activation (66). 15

16

#### 17 CONCLUSION

An ELSD that is able to bridge patients with liver failure either to recovery or to the state they 18 19 were in, prior to the present deterioration, remains an unmet medical need. The main impediments to the development of an effective device can be thought of as being either 20 patient related or device related. It is clear that once multiorgan failure is established, it is 21 probably too late for an ELSD to be effective: in this situation the sole aim of ELSD treatment 22 should be a bridge to transplant. Therefore, clinical trials need to include patients at risk of 23 progression to multiorgan failure. The number of patients that will be required to attain 24 adequate power will be high. It is also clear that the currently available devices show 25 improvements in pathophysiological variables known to be associated with liver failure, but 26 27 only one, TPE, has demonstrated survival benefit. The deficiencies of the currently available 28 devices have inspired the newer devices, which are currently in clinical trials or due to enter

1 trials shortly. As ACLF has now been defined and the pathophysiology of both ALF and ACLF becomes clearer, it is very likely that an effective ELSD will emerge. Moreover further 2 3 indications for ELSD may become evident. Indeed, new opportunity has arisen following the discovery of the new directly acting anti-viral drugs for Hepatitis C virus infection, which have 4 5 been shown to reverse the severity of cirrhosis in many patients (68). One can envisage a 6 situation whereby, Hepatitis C patients with decompensated cirrhosis are treated with ELSDs 7 as out-patients for weeks and months, while the new directly acting anti-viral drugs take 8 effect, negating the need for liver transplantation.

9

### 10 **REFERENCES**

11 1. Jalan R, Yurdaydin C, Bajaj JS, Acharya SK, Arroyo V, Lin HC, et al. Toward an

improved definition of acute-on-chronic liver failure. Gastroenterology 2014;147:4-10.

Jalan R, Sen S, Williams R. Prospects for extracorporeal liver support. Gut
 2004;53:890-8.

Antoniades CG, Berry PA, Wendon JA, Vergani D. The importance of immune
 dysfunction in determining outcome in acute liver failure. J Hepatol 2008;49:845-61.

17 4. O'Grady JG, Williams R. Classification of acute liver failure. Lancet 1993;342:743.

Jalan R, Gines P, Olson JC, Mookerjee RP, Moreau R, Garcia-Tsao G, et al. Acuteon chronic liver failure. J Hepatol 2012;57:1336-48.

Moreau R, Jalan R, Gines P, Pavesi M, Angeli P, Cordoba J, et al. Acute-on-chronic
 liver failure is a distinct syndrome that develops in patients with acute decompensation of
 cirrhosis. Gastroenterology 2013;144:1426-37, 37 e1-9.

7. Jalan R, Saliba F, Pavesi M, Amoros A, Moreau R, Gines P, et al. Development and
validation of a prognostic score to predict mortality in patients with acute-on-chronic liver
failure. J Hepatol 2014;61:1038-47.

Jalan R, Pavesi M, Saliba F, Amoros A, Fernandez J, Holland-Fischer P, et al. The
 CLIF Consortium Acute Decompensation score (CLIF-C ADs) for prognosis of hospitalised
 cirrhotic patients without acute-on-chronic liver failure. J Hepatol 2015;62:831-40.

1	9.	Arroyo V, Moreau R, Jalan R, Gines P. Acute-on-chronic liver failure: A new
2	syndro	ome that will re-classify cirrhosis. J Hepatol 2015;62:S131-43.
3	10.	Rozga J. Liver support technologyan update. Xenotransplantation 2006;13:380-9.
4	11.	Stange J, Mitzner S, Ramlow W, Gliesche T, Hickstein H, Schmidt R. A new
5	proced	dure for the removal of protein bound drugs and toxins. ASAIO J 1993;39:M621-5.
6	12.	Mitzner SR, Stange J, Klammt S, Risler T, Erley CM, Bader BD, et al. Improvement
7	of hep	atorenal syndrome with extracorporeal albumin dialysis MARS: results of a
8	prospe	ective, randomized, controlled clinical trial. Liver Transpl 2000;6:277-86.
9	13.	Stange J, Ramlow W, Mitzner S, Schmidt R, Klinkmann H. Dialysis against a
10	recycle	ed albumin solution enables the removal of albumin-bound toxins. Artif Organs
11	1993;′	17:809-13.
12	14.	Heemann U, Treichel U, Loock J, Philipp T, Gerken G, Malago M, et al. Albumin
13	dialysi	s in cirrhosis with superimposed acute liver injury: a prospective, controlled study.
14	Hepat	ology 2002;36:949-58.
15	15.	Falkenhagen D, Strobl W, Vogt G, Schrefl A, Linsberger I, Gerner FJ, et al.
16	Fractio	onated plasma separation and adsorption system: a novel system for blood purification
17	to rem	ove albumin bound substances. Artif Organs 1999;23:81-6.
18	16.	Rifai K, Ernst T, Kretschmer U, Bahr MJ, Schneider A, Hafer C, et al. Prometheusa
19	new e	xtracorporeal system for the treatment of liver failure. J Hepatol 2003;39:984-90.
20	17.	Peszynski P, Klammt S, Peters E, Mitzner S, Stange J, Schmidt R. Albumin dialysis:
21	single	pass vs. recirculation (MARS). Liver 2002;22 Suppl 2:40-2.
22	18.	Sauer IM, Goetz M, Steffen I, Walter G, Kehr DC, Schwartlander R, et al. In vitro
23	compa	arison of the molecular adsorbent recirculation system (MARS) and single-pass
24	album	in dialysis (SPAD). Hepatology 2004;39:1408-14.
25	19.	Stenbog P, Busk T, Larsen FS. Efficacy of liver assisting in patients with hepatic
26	encep	halopathy with special focus on plasma exchange. Metab Brain Dis 2013;28:333-5.

Stadlbauer V, Krisper P, Beuers U, Haditsch B, Schneditz D, Jung A, et al. Removal
 of bile acids by two different extracorporeal liver support systems in acute-on-chronic liver
 failure. ASAIO J 2007;53:187-93.

4 21. Krisper P, Haditsch B, Stauber R, Jung A, Stadlbauer V, Trauner M, et al. In vivo
5 quantification of liver dialysis: comparison of albumin dialysis and fractionated plasma
6 separation. J Hepatol 2005;43:451-7.

Khuroo MS, Farahat KL. Molecular adsorbent recirculating system for acute and
acute-on-chronic liver failure: a meta-analysis. Liver Transpl 2004;10:1099-106.

9 23. El Banayosy A, Kizner L, Schueler V, Bergmeier S, Cobaugh D, Koerfer R. First use
10 of the Molecular Adsorbent Recirculating System technique on patients with hypoxic liver
11 failure after cardiogenic shock. ASAIO J 2004;50:332-7.

12 24. Mitzner SR, Klammt S, Peszynski P, Hickstein H, Korten G, Stange J, et al.

13 Improvement of multiple organ functions in hepatorenal syndrome during albumin dialysis

14 with the molecular adsorbent recirculating system. Ther Apher 2001;5:417-22.

15 25. Schmidt LE, Wang LP, Hansen BA, Larsen FS. Systemic hemodynamic effects of

16 treatment with the molecular adsorbents recirculating system in patients with hyperacute

17 liver failure: a prospective controlled trial. Liver Transpl 2003;9:290-7.

18 26. Chen S, Zhang L, Shi Y, Yang X, Wang M. Molecular Adsorbent Recirculating

19 System: clinical experience in patients with liver failure based on hepatitis B in China. Liver

20 2002;22 Suppl 2:48-51.

21 27. Hessel FP, Mitzner SR, Rief J, Guellstorff B, Steiner S, Wasem J. Economic

22 evaluation and 1-year survival analysis of MARS in patients with alcoholic liver disease.

23 Liver Int 2003;23 Suppl 3:66-72.

24 28. Sen S, Davies NA, Mookerjee RP, Cheshire LM, Hodges SJ, Williams R, et al.

25 Pathophysiological effects of albumin dialysis in acute-on-chronic liver failure: a randomized

controlled study. Liver Transpl 2004;10:1109-19.

Catalina MV, Barrio J, Anaya F, Salcedo M, Rincon D, Clemente G, et al. Hepatic
 and systemic haemodynamic changes after MARS in patients with acute on chronic liver
 failure. Liver Int 2003;23 Suppl 3:39-43.

30. Sen S, Mookerjee RP, Cheshire LM, Davies NA, Williams R, Jalan R. Albumin
dialysis reduces portal pressure acutely in patients with severe alcoholic hepatitis. J Hepatol
2005;43:142-8.

31. Laleman W, Wilmer A, Evenepoel P, Elst IV, Zeegers M, Zaman Z, et al. Effect of the
molecular adsorbent recirculating system and Prometheus devices on systemic

9 haemodynamics and vasoactive agents in patients with acute-on-chronic alcoholic liver

10 failure. Crit Care 2006;10:R108.

11 32. Hassanein TI, Tofteng F, Brown RS, Jr., McGuire B, Lynch P, Mehta R, et al.

12 Randomized controlled study of extracorporeal albumin dialysis for hepatic encephalopathy

in advanced cirrhosis. Hepatology 2007;46:1853-62.

14 33. Banares R, Nevens F, Larsen FS, Jalan R, Albillos A, Dollinger M, et al.

15 Extracorporeal albumin dialysis with the molecular adsorbent recirculating system in acute-

16 on-chronic liver failure: the RELIEF trial. Hepatology 2013;57:1153-62.

17 34. Saliba F, Camus C, Durand F, Mathurin P, Letierce A, Delafosse B, et al. Albumin

dialysis with a noncell artificial liver support device in patients with acute liver failure: a

19 randomized, controlled trial. Ann Intern Med 2013;159:522-31.

20 35. Olin P, Hausken J, Foss A, Karlsen TH, Melum E, Haugaa H. Continuous molecular

adsorbent recirculating system treatment in 69 patients listed for liver transplantation. Scand

22 J Gastroenterol 2015;50:1127-34.

23 36. Lexmond WS, Van Dael CM, Scheenstra R, Goorhuis JF, Sieders E, Verkade HJ, et

al. Experience with molecular adsorbent recirculating system treatment in 20 children listed

for high-urgency liver transplantation. Liver Transpl 2015;21:369-80.

26 37. Cisneros-Garza LE, Munoz-Ramirez Mdel R, Munoz-Espinoza LE, Ruiz Velasco JA,

27 Moreno-Alcantar R, Marin-Lopez E, et al. The molecular adsorbent recirculating system as a

liver support system: summary of Mexican experience. Ann Hepatol 2014;13:240-7.

Marangoni R, Bellati G, Castelli A, Romagnoli E. Development of high-efficiency
 molecular adsorbent recirculating system: preliminary report. Artif Organs 2014;38:879-83.
 39. Evenepoel P, Laleman W, Wilmer A, Claes K, Maes B, Kuypers D, et al. Detoxifying
 capacity and kinetics of prometheus--a new extracorporeal system for the treatment of liver
 failure. Blood Purif 2005;23:349-58.

40. Nyckowski P, Skwarek A, Zieniewicz K, Patkowski W, Alsharabi A, Wroblewski T, et
al. Orthotopic liver transplantation for fulminant hepatic failure. Transplant Proc 2006;38:21920.

9 41. Meijers BK, Verhamme P, Nevens F, Hoylaerts MF, Bammens B, Wilmer A, et al.

Major coagulation disturbances during fractionated plasma separation and adsorption. Am J
 Transplant 2007;7:2195-9.

42. Senturk E, Esen F, Ozcan PE, Rifai K, Pinarbasi B, Cakar N, et al. The treatment of
acute liver failure with fractionated plasma separation and adsorption system: Experience in
85 applications. J Clin Apher 2010;25:195-201.

43. Kribben A, Gerken G, Haag S, Herget-Rosenthal S, Treichel U, Betz C, et al. Effects
of fractionated plasma separation and adsorption on survival in patients with acute-on-

17 chronic liver failure. Gastroenterology 2012;142:782-9.

44. Kortgen A, Rauchfuss F, Gotz M, Settmacher U, Bauer M, Sponholz C. Albumin
dialysis in liver failure: comparison of molecular adsorbent recirculating system and single
pass albumin dialysis--a retrospective analysis. Ther Apher Dial 2009;13:419-25.

45. Karvellas CJ, Bagshaw SM, McDermid RC, Stollery DE, Bain VG, Gibney RT. A
case-control study of single-pass albumin dialysis for acetaminophen-induced acute liver
failure. Blood Purif 2009;28:151-8.

46. Umgelter A, Lange K, Kornberg A, Buchler P, Friess H, Schmid RM. Orthotopic liver
transplantation in critically ill cirrhotic patients with multi-organ failure: a single-center

26 experience. Transplant Proc 2011;43:3762-8.

47. Larsen FS, Schmidt LE, Bernsmeier C, Rasmussen A, Isoniemi H, Patel VC, et al.
 High-volume plasma exchange in patients with acute liver failure: An open randomised
 controlled trial. J Hepatol 2015;Aug 29. pii:S0168-8278(15)00590-5.

4 48. van Wenum M, Chamuleau RA, van Gulik TM, Siliakus A, Seppen J, Hoekstra R.
5 Bioartificial livers in vitro and in vivo: tailoring biocomponents to the expanding variety of
6 applications. Expert Opin Biol Ther 2014;14:1745-60.

7 49. Zhou N, Li J, Zhang Y, Lu J, Chen E, Du W, et al. Efficacy of coupled low-volume
8 plasma exchange with plasma filtration adsorption in treating pigs with acute liver failure: A
9 randomised study. J Hepatol 2015;63:378-87.

10 50. Sussman NL, Chong MG, Koussayer T, He DE, Shang TA, Whisennand HH, et al.

11 Reversal of fulminant hepatic failure using an extracorporeal liver assist device. Hepatology
12 1992;16:60-5.

13 51. Sussman NL, Gislason GT, Conlin CA, Kelly JH. The Hepatix extracorporeal liver
14 assist device: initial clinical experience. Artif Organs 1994;18:390-6.

15 52. Ellis AJ, Hughes RD, Wendon JA, Dunne J, Langley PG, Kelly JH, et al. Pilot-

16 controlled trial of the extracorporeal liver assist device in acute liver failure. Hepatology

17 1996;24:1446-51.

18 53. Vital. Therapies Inc. Vital Therapies® targeting liver disease.

19 <u>http://vitaltherapies.com/</u>. Accessed July 2015.

20 54. Nyberg SL, Remmel RP, Mann HJ, Peshwa MV, Hu WS, Cerra FB. Primary

21 hepatocytes outperform Hep G2 cells as the source of biotransformation functions in a

bioartificial liver. Ann Surg 1994;220:59-67.

23 55. Duan Z-P, Zhang J, Xin S, Chen JM, He D, Brotherton JD, et al. Interim results of

randomized controlled trial of ELAD<sup>™</sup> in acute and chronic liver disease. Hepatology

25 2007;46:247A.

26 56. US. National Institutes of Health. ClinicalTrials.gov.

27 https://www.clinicaltrials.gov/ct2/results?term=ELAD%E2%84%A2+liver&no\_unk=Y.

Accessed July 2015.

57. Vital. Therapies Inc. Vital Therapies announces that topline results of VTI-208 fail to
 achieve primary or secondary endpoints of improvement in overall survival pre-specified
 exploratory subset analyses suggest efficacy trends.

4 <u>http://ir.vitaltherapies.com/releasedetail.cfm?ReleaseID=928472</u>. Accessed August 2015.

5 58. van de Kerkhove MP, Poyck PP, Deurholt T, Hoekstra R, Chamuleau RA, van Gulik
6 TM. Liver support therapy: an overview of the AMC-bioartificial liver research. Dig Surg
7 2005;22:254-64.

8 59. Nibourg GA, Hoekstra R, van der Hoeven TV, Ackermans MT, Hakvoort TB, van
9 Gulik TM, et al. Increased hepatic functionality of the human hepatoma cell line HepaRG
10 cultured in the AMC bioreactor. Int J Biochem Cell Biol 2013;45:1860-8.

11 60. Nibourg GA, Chamuleau RA, van der Hoeven TV, Maas MA, Ruiter AF, Lamers WH,

12 et al. Liver progenitor cell line HepaRG differentiated in a bioartificial liver effectively supplies

liver support to rats with acute liver failure. PLoS One 2012;7:e38778.

14 61. Nyberg SL, Hardin J, Amiot B, Argikar UA, Remmel RP, Rinaldo P. Rapid, large-

15 scale formation of porcine hepatocyte spheroids in a novel spheroid reservoir bioartificial

16 liver. Liver Transpl 2005;11:901-10.

17 62. McIntosh MB, Corner SM, Amiot BP, Nyberg SL. Engineering analysis and

development of the spheroid reservoir bioartificial liver. Conf Proc IEEE Eng Med Biol Soc
2009;2009:5985-8.

63. Glorioso JM, Mao SA, Rodysill B, Mounajjed T, Kremers WK, Elgilani F, et al. Pivotal
preclinical trial of the spheroid reservoir bioartificial liver. J Hepatol 2015;63:388-98.

22 64. Al-Chalabi A, Matevossian E, AK VT, Luppa P, Neiss A, Schuster T, et al. Evaluation

23 of the Hepa Wash(R) treatment in pigs with acute liver failure. BMC Gastroenterol

24 2013;13:83.

25 65. Evenepoel P, Laleman W, Wilmer A, Claes K, Kuypers D, Bammens B, et al.

26 Prometheus versus molecular adsorbents recirculating system: comparison of efficiency in

two different liver detoxification devices. Artif Organs 2006;30:276-84.

1 (	66.	Lee KC, Baker LA, Stanzani G, Alibhai H, Chang YM, Jimenez Palacios C, et al.
-----	-----	---

2 Extracorporeal liver assist device to exchange albumin and remove endotoxin in acute liver

- 3 failure: Results of a pivotal pre-clinical study. J Hepatol 2015;63:634-42.
- 4 67. Morgera S, Haase M, Rocktaschel J, Bohler T, Vargas-Hein O, Melzer C, et al.
- 5 Intermittent high-permeability hemofiltration modulates inflammatory response in septic
- 6 patients with multiorgan failure. Nephron Clin Pract 2003;94:c75-80.
- 7 68. Pawlotsky JM, Feld JJ, Zeuzem S, Hoofnagle JH. From non-A, non-B hepatitis to
- 8 hepatitis C virus cure. J Hepatol 2015;62:S87-99.
- 9

# 1 Table 1: Types of Liver Failure

	Hyperacute/	Sub acute	ACLF	Decompensated
	Acute		underlying	cirrhosis
			cirrhosis	
Time from	Weeks	Months	Weeks	Years
symptoms to				
failure				
Common	Toxic	?Viral	Variable	Variable
aetiology				
Precipitating	Liver injury	Liver Injury	Infection	Unknown
event			Alcohol	Infection (others)
			Unknown	
Prognostic	Kings	Kings	CLIF C score	MELD
score				
Potential for	High	Poor	Unknown	Poor
regeneration				

2

3 Adapted from Jalan et al. Gastroenterology 2014 (1).

- 1 Table 2: The CLIF Consortium organ failure score for the diagnosis of acute on
- 2 chronic liver failure

Organ System	Score = 1	Score = 2	Score = 3
Liver (mg/dl)	Bilirubin < 6	6 ≤ Bilirubin ≤ 12	Bilirubin >12
Kidney (mg/dl)	Creatinine <2.0	Creatinine ≥2.0 or	Creatinine ≥3.5 or
		<3.5	renal replacement
Brain	Grade 0	Grade 1-2	Grade 3-4
(West-Haven)			
Coagulation	INR < 2.0	2.0 ≤ INR < 2.5	INR ≥ 2.5
Circulation	MAP ≥70 mm/Hg	MAP <70 mm/Hg	Vasopressors
Respiratory:	>300	≤300 - > 200	≤200
PaO <sub>2</sub> /FiO <sub>2</sub>	>357	>214- ≤357	≤214
or SpO <sub>2</sub> /FiO <sub>2</sub>			

3 No ACLF: Patients with no organ failure; patients with single hepatic, coagulation, circulation

4 or respiratory failure, serum creatinine <1.5 mg/dl and no HE; or patient with cerebral failure

5 and serum creatinine <1.5 mg/dl.

6 ACLF 1: Patients with renal failure or patients with other single organ failure with either

serum creatinine  $\geq$  1.5 and < 2 mg/dl and/or HE grade 1-2.

8 ACLF 2: Patients with 2 organ failures.

9 ACLF 3: Patients with 3 or more organ failures.

10 Adapted from Jalan et al. Journal of Hepatology 2014 (7)

11

## 1 Table 3: Reported human randomised controlled clinical trials for ELSDs with survival as the primary outcome measure.

2 Data from intention to treat (ITT) and per protocol (PP) analyses are included where reported separately. (ALF, acute liver failure; ACLF, acute-

3 on-chronic liver failure; n.s., no significant difference between groups; SMT, standard medical therapy; HVP, high-volume therapeutic plasma

4 exchange)

Liver support	Study name	Type of trial	Patient	Number of patients	Primary outcome	Secondary outcomes	Safety profile
device	or identifier		type	randomised		(only significant	
				(patients excluded		outcomes described)	
				after randomisation			
				given in brackets)			
MARS®	The RELIEF	Multi-centre	ACLF	Total=189	28-day ITT survival: MARS <sup>®</sup> , 61%; SMT,	At day 4, MARS®	Incidence of
	Trial (33)	open		MARS <sup>®</sup> =95	59% (n.s.).	resulted in a significant	severe adverse
		randomised		SMT=94	28-day PP survival: MARS <sup>®</sup> , 60%; SMT,	reduction in serum	events was
		controlled		(-ITT analysis: 5	59% (n.s.).	creatinine, bilirubin and	similar in MARS®
		trial		exclusions per group		hepatic encephalopathy	and SMT groups
				-PP analysis: 24		scores compared to	
				MARS <sup>®</sup> and 9 SMT		SMT.	
				exclusions)			

MARS®	The	Multi-centre	ALF	Total=110	6-month ITT survival: MARS <sup>®</sup> , 85%;		Incidence of
	FULMAR	open		MARS <sup>®</sup> =57	SMT, 76% (n.s.).		severe adverse
	Trial (34)	randomised		SMT=53	6-month PP survival: MARS <sup>®</sup> , 82%; SMT,		events was
		controlled		(-ITT analysis: 4	76% (n.s.).		similar in MARS®
		trial		exclusions per group			and SMT groups
				-PP analysis: 18			
				MARS <sup>®</sup> and 4 SMT			
				exclusions)			
Prometheus®	The HELIOS	Multi-centre	ACLF	Total=145	28-day ITT survival: Prometheus <sup>®</sup> , 66%;	At day 28,	Incidence of
	trial (43)	open		Prometheus <sup>®</sup> =77	SMT, 63% (n.s.).	Prometheus <sup>®</sup> resulted	severe adverse
		randomised		SMT=68	28-day PP survival: Prometheus <sup>®</sup> , 71%;	in a significant	events was
		controlled		(-ITT analysis: 0	SMT, 67% (n.s.).	reduction in serum	similar in
		trial		exclusions	90-day ITT survival: Prometheus <sup>®</sup> , 47%;	bilirubin compared to	Prometheus®
				-PP analysis: 22	SMT, 38% (n.s.).	SMT.	and SMT groups
				Prometheus <sup>®</sup> and 14	90-day PP survival: Prometheus <sup>®</sup> , 41%;		
				SMT exclusions)	SMT, 39% (n.s.).		
					(Figure 2)		

High-volume	ClinicalTrials	Multi-centre	ALF	Total=183	Survival to hospital discharge: HVP, 59%;	On day 1 to day 7, HVP	Incidence of
therapeutic	.gov number	open		HVP=92	SMT, 48% (P=0.008).	resulted in significant	severe adverse
plasma	NCT002247	randomised		SMT=91	(Figure 3)	reduction in	events was
exchange	05 (47)	controlled		(1 SMT excluded		international	similar in HVP
(HVP)		trial		after randomisation)		normalised ratio,	and SMT groups
						bilirubin, ALT, SOFA-	
						score and CLIF-score.	
ELAD™	(55)	Multi-centre	Chronic	Total=60	30-day ITT transplant-free survival:		ELAD™ was
		open	hepatitis	ELAD™=40	ELAD™, 80%; SMT, 50% (P=0.03).		associated with
		randomised	B or C	SMT=20	30-day PP transplant-free survival:		significant
		controlled	with	(-ITT analysis: 0	ELAD™, 86%; SMT, 47% (P=0.004).		thrombocytopeni
		trial	acute	exclusions			a, whilst SMT
			decomp-	-PP analysis: 5			was not.
			ensation	ELAD™ and 1 SMT			
				exclusions)			

## 1 FIGURE LEGENDS

## 2 Figure 1: Survival data from the RELIEF trial.

28-day survival for MARS<sup>®</sup> (light grey line) compared to standard medical therapy, SMT
(dark grey line) with intention to treat analysis on the left and per protocol analysis on the
right. Number of survivors at each time point is inserted into the graphs. See Table 3 for
study details. (Reproduced with permission from Hepatology by John Wiley and Sons (33))

## 8 Figure 2: Survival data from the HELIOS trial.

9 90-day intention to treat survival for Prometheus<sup>®</sup>, FPSA+SMT, compared to standard

10 medical therapy, SMT. See Table 3 for study details. (Reproduced with permission from

- 11 Gastroenterology by Elsevier (43))
- 12

## 13 Figure 3: Survival data from the high-volume plasma exchange trial.

14 90-day intention to treat survival for high-volume plasma exchange, HVP, compared to

- 15 standard medical therapy, SMT. See Table 3 for study details. (Reproduced with permission
- 16 from Journal of Hepatology by Elsevier (47))
- 17

# Figure 1









